Streptococcus penumoniae meningitis

Clinical and experimental studies

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1. INTRODUCTION

Before the introduction of antibiotics (sulphonamides in the 1930's and penicillin 1940's), meningitis due to *Streptococcus pneumoniae* ended without exception in the death of the patients (Netter 1887; Southard and Keene 1906). Several desperate therapeutic attempts such as drainage of cerebrospinal fluid and treatment with optochin, bile salt, or pneumococcal antiserum were performed on experimental basis during the pre-antibiotic period, but without clinical success (Kolmer 1929). Although treatment with antibiotics made *S. pneumoniae* meningitis a curable disease (Appelbaum and Nelson 1945; Finland et al 1938), the morbidity and mortality from the disease have not changed significantly over decades (**Figure 1**) and remain unacceptably high, despite continuous improvements in intensive care technology and the introduction of new more potent antibiotics (Swartz 2004).

The exact mechanism leading to the devastating outcome of *S. pneumoniae* meningitis is not fully elucidated but may include a direct harmful effect of the pathogen itself and a host immune reaction against the invading pathogen that continues to evolve after the bacteria are killed from antibiotic therapy. Therefore, therapeutic intervention should not only be directed against the invading pathogen, but also against the harmful effects of the host immune response. This has led to an increasing interest in studying the patho-



Figure 1. Mortality of pneumococcal meningitis over decades. The figure is reproduced according to (Swartz 2004) and includes case fatality rates from Danish studies.

genesis and pathophysiology of bacterial meningitis and to a search for new adjunctive therapeutic strategies to improve the outcome of the disease (for a review: (Koedel et al 2002a; Meli et al 2002; Nau and Bruck 2002)).

Although therapeutic intervention - from the clinician's perspective - predominantly should target pathological events occurring late during the course of meningitis, studies of all aspects of the disease may not alone contribute to an increasing knowledge of pathogenesis and pathophysiology of bacterial meningitis, but also to a better outcome of the disease. Indeed, further progress may rather come from prevention of pneumococcal meningitis as has been the case with Haemophilus influenzae meningitis and from early identification of risk factors and predisposing conditions than from improvements in the treatment regimens (Swartz 2004). Strikingly, most therapeutic interventions used for more than a half century in the treatment of patients with bacterial meningitis (e.g. agents, dose and duration of antibiotic therapy (Prasad et al 2004), fluid restriction (Møller et al 2001a; Oates-Whitehead et al 2005), osmotic therapy (Nau 2000), hyperventilation (Ashwal et al 1994)) are not based on randomised clinical trials and are still controversial (Tunkel et al 2004), and also recent results of the efficacy of adjunctive therapy with dexamethasone are conflicting (de Gans and van de Beek 2002; Molyneux et al 2002). Therefore, better documentation and new treatment options are still warranted.

The clinical meningitis studies generated until now can primarily be grouped as descriptive and intervention studies. The descriptive studies are most frequent in numbers and include: 1) characterisation of the bacterial aetiology, 2) clinical characteristics including epidemiological data and outcome data, 3) non-invasive measurements (e.g. MR, Laser-Doppler flowmetry), 4) CSF and blood analysis for the evaluation of antibiotic pharmacokinetics, characterisation of pathophysiological mediators and their diagnostic- and prognostic use, and 5) autopsy studies (histopathology and immunohistochemistry). The intervention studies include the treatment efficacy of 1) antibiotics, 2) adjunctive therapy with corticosteroids, and 3) other kind of therapy (e.g. hyperventilation, fluid restriction, osmotic therapy) and are few in numbers, because it demands considerable resources during several years to perform multicentre randomised clinical trials. In addition, there are obvious limitations in clinical meningitis studies due to lack of possibilities for invasive sampling procedures during the disease. Therefore, the use of animal models is essential for a better understanding of the pathogenesis and pathophysiology of pneumococcal meningitis.

Aim of own studies. In continuation of the work presented in the Ph.D. thesis "The inflammatory response in bacterial meningitis. An experimental meningitis model" (Østergaard 2000), which was obtained in the rabbit meningitis model, we wanted to further address various aspects of the pathogenesis and pathophysiology of bacterial meningitis with special focus on pneumococcal meningitis. When appropriate, we wanted to test our research goals and hypothesis using clinical and experimental studies. We wanted to study the following issues:

- 1. Evaluation of clinical features and prognostic factors (VIII, IX, X, XI).
- 2. CSF evaluation for the identification of new diagnostic and prognostic tools (I, VI, VII).
- 3. Pharmacokinetic and pharmacodynamic study of potential new antibiotics in the treatment of meningitis (II).
- 4. A further characterisation of the mechanism behind the meningeal inflammatory response (III, IV, V, VIII, XI).
- 5. A characterisation of histopathological alterations (VIII).

2. EPIDEMIOLOGY

In Denmark, pneumococcal meningitis accounts for approximately 100 cases per year (incidence of $\sim 2/100,000$ cases per year) (Østergaard et al 2005), which is 10-1000 times lower than the incidence of



Figure 2. Age distribution and mortality of pneumococcal meningitis in Denmark in 1999-2000.

other invasive pneumococcal diseases (e.g. bacteraemia: ~20/100.000 cases per year (Konradsen and Kaltoft 2002), pneumonia: ~300/ 100.000 cases per year (Austrian 1981b)). The incidence of pneumo-coccal meningitis seems not to have varied significantly over decades in Denmark (Kaltoft et al 2000; Konradsen and Kaltoft 2002; Lund 1970; Nielsen and Henrichsen 1993; Østergaard et al 2005; Pedersen and Henrichsen 1983) or to differ significantly between industrilised countries (Eriksson et al 2000; Lexau et al 2005; Rendi-Wagner et al 2004; Weisfelt et al 2006), whereas exact epidemiological data from developing countries still are lacking (Gordon et al 2000).

The incidence of pneumococcal meningitis is highest during the winter season and varies according to age groups with more cases in children under two years of age (~25% of total cases) and in adults with increasing age (Figure 2, (Kaltoft et al 2000; Østergaard et al 2005)). The use of paediatric conjugate vaccine or polysaccharide vaccines to adults may change the epidemiology of meningitis, because the use of pneumococcal vaccines has shown a beneficial effect by reducing the risk of predisposing conditions to pneumococcal meningitis such as bacteraemia (Cutts et al 2005; Jackson et al 2003; Lexau et al 2005), and otitis media (Eskola et al 2001; Prymula et al 2006) and nasopharyngeal carriage (Dagan et al 2002). Indeed, a reduced transmission to non-vaccinated groups (herd immunity) seems to be an important issue for preventing invasive pneumococcal disease (Kyaw et al 2006; McIntosh et al 2005). However, serotype replacement that has been observed after the introduction of pneumococcal conjugate vaccine (Lexau et al 2005) is a cause of concern and should be monitored carefully in future studies.

In Denmark less than 10% of CSF isolates have reduced susceptibility to penicillin (Østergaard et al 2005), whereas high prevalence of penicillin – and cephalosporin resistance has emerged in countries like Australia, Spain, South Africa and USA (Whitney et al 2000). Importantly, the use of pneumococcal vaccines also had an effect by reducing the risk of invasive disease caused by resistant strains (Kyaw et al 2006).

3. CLINICAL PRESENTATION

Predisposing condition for developing pneumococcal meningitis is besides leak of the blood/brain barrier, which is found in ~11% of cases (Østergaard et al 2005) (e.g. oto – and rhinorrhoae after basilar skull fractures (Ratilal et al 2006), cochlear implants (Reefhuis et al 2003)), an increased susceptibility of the host for invasive pneumococcal disease. This includes an impaired immune response against pneumococcal infection (e.g. asplenia, various immune deficiencies, use of immunosuppressing therapy (Fraser et al 1973)), but also recurrent otitis media and day care attendance have been found to dispose for invasive pneumococal disease (Takala et al 1995). We found that approximately 1 of every 3 cases with pneumococcal meningitis are secondary to an otogenic focus, whereas $\sim 20\%$ are due to a lung focus, $\sim 8\%$ to a sinusitic focus and in $\sim 40\%$ no primary infection focus can be found (Østergaard et al 2005), results which have persistently been observed over decades (Bohr et al 1985; Geiseler et al 1980). The presence of an accompanying focus depended on age groups with a lung focus more frequently observed among adult cases than among children (26% vs. 7%, respectively), whereas a higher proportion of children had no primary infection focus (64% vs. 35% in adult cases). This could reflect the higher prevalence of nasopharyngeal carriage in children than in adults (Leino et al 2001). However, an otogenic focus was as frequent in adults as in children, despite that the incidence of otitis media most likely is significantly higher among children than among adults. Our results also suggested that inadequate antibiotic therapy of otitis media could be a risk factor for developing pneumococcal meningitis (Østergaard et al 2006a).

The classical signs of meningitis (fever, back rigidity, decreased consciousness, and convulsion) are also characteristic clinical findings in pneumococcal meningitis and was found in 96%, 55%, 94%, and 12% of cases, respectively (Østergaard et al 2005), which is in accordance with recent studies in children (Casado-Flores et al 2005) and adults (Weisfelt et al 2006). Other clinical findings on admission include headache (~70% of adults), paresis (~10%), cranial nerve palsies (~10%), papilloedema (~5%), and tense fontanel (~50-70% of young children) (Casado-Flores et al 2005; Weisfelt et al 2006). Since back rigidity was less frequently observed than fever and a decreased consciousness, which are clinical features found in patients with other diseases than meningitis (e.g. sepsis), it might result in differential diagnostic difficulties. Indeed, not all lumbar punctures were performed on admission, and we found that a diagnostic CT-scan was performed in ~10% of cases before lumbar puncture, which delayed the initiation of antibiotic therapy (Østergaard et al 2005). Strikingly, ~57% of Dutch adult cases had a CTscan before lumbar puncture (Weisfelt et al 2006), which could reflect difference in treatment protocols between countries. Indeed, the majority of Danish adult cases were admitted to internal medicine departments in contrast to The Netherlands, where adult meningitis patients were admitted to neurological departments.

4. CSF EVALUATION

Diagnostic use. Routine examination of the CSF for bacteria, WBC including differential counts, and concentrations of glucose and protein is the primary investigation to diagnose meningitis. Pneumococci can be visualized in Gram staining in ~90% and can be grown from the CSF in 98% of documented cases with pneumococcal meningitis, and in addition a positive blood culture is observed in ~67% of cases (Gray and Fedorko 1992; Østergaard et al 2005). However, in ~40% of a patient cohort with purulent meningitis admitted to a Danish hospital, the causative pathogen was not detected (Østergaard et al 2002b; Østergaard et al 2004a). Improvements in molecular techniques have emerged in the microbiological laboratory during the last two decades, but until now polymerase chain reaction (PCR) techniques have not improved the sensitivity in the detection of CSF bacterial pathogens (Corless et al 2001), in contrast to the significant improvements that have been obtained in the determination of viral pathogens (Koskiniemi et al 2001). Collection of the CSF in enriched culture media for transport to the microbiological laboratory has been shown to augment the determination of the bacterial pathogen (Lessing and Bowler 1996), and in cases receiving antibiotics before the CSF tap, PCR techniques may have a beneficial role (Cherian et al 1998).

In pneumococcal meningitis, we found that WBC counts varied

between 1-2 cells/ μ L to up to 50.000 cells/ μ L (Østergaard et al 2005), whereas the elevation in CSF protein levels and the decrease in CSF glucose levels often were more pronounced in pneumococcal meningitis than in other forms of community-acquired bacterial meningitis including *Neisseria meningitidis* meningitis (Østergaard et al 2002b; Østergaard et al 2004a), however, the results of CSF biochemical analysis are not always conclusive to distinguish bacterial from viral meningitis. Therefore, clinical studies of other CSF candidates (see **Table 1** for a selected number of studies) may be useful for differential diagnostic purposes.

We studied IL-8 in meningitis, because it has a key role in neutrophil chemotaxis and recruitment in vitro and in vivo (Harada et al 1994; Smith et al 1991). CSF IL-8 levels were highly elevated en meningitis and were to some degree useful in distinguishing between bacterial and viral meningitis with a sensitivity, specificity and the positive predictive value of ~81%, ~92%, and ~96%, respectively (Østergaard et al 1996). CSF levels of TNF α and IL-1 β (Lopez-Cortes et al 1993; Ramilo et al 1990a) and serum levels of CRP (Roine et al 1992) and PCT (Schwarz et al 2000) may also help to distinguish between bacterial and viral meningitis.

Prognostic value. A high number of bacteria in the CSF or a high CSF concentration of bacterial antigens (Feldman 1977; Mertsola et al 1991) as well as a high CSF concentration of the pneumococcal degradation product, lipoteichoic acid have been correlated with a poor outcome of meningitis (Schneider et al 1999). In pneumococcal meningitis, we found a significant association between poor outcome and alterations in routine CSF analysis (e.g. low number of CSF WBC, low CSF/blood glucose ratio, high CSF protein levels) (Østergaard et al 2005), which is in accordance with some studies (Kastenbauer and Pfister 2003; Weisfelt et al 2006), whereas others have not been able to show such an association (Bohr et al 1985).

Also, we studied YKL-40, a chitinase-related protein, in meningitis, because it had a prognostic value in other diseases such as pneumococcal bacteraemia and cancer (Cintin et al 1999; Kronborg et al 2002). We found that CSF YKL-40 levels were elevated in meningitis – in particular in patients with pneumococcal meningitis – and were related to the severity of the infection (Østergaard et al 2002b). We also studied suPAR, the soluble form of the urokinasetype plasminogen activator receptor, which is involved in proteolysis of the basement membrane and leukocyte migration (Blasi and Carmeliet 2002), because it had a prognostic value in pneumococcal bacteraemia and cancer (Stephens et al 1999; Wittenhagen et al 2004). We found that CSF levels of suPAR were elevated in bacterial meningitis and were correlated to a poor outcome (Østergaard et al 2004a).

In conclusion, clinical meningitis studies of various CSF parameters have contributed to a better understanding of the meningeal inflammatory process. However, several limitations in clinical studies of CSF parameters exist: 1) Patients may have individual immune response, 2) most studies include few and poorly characterised patients, and 3) variation in sampling time occurs during the course of meningitis. Thus, despite that CSF mediators to some degree were useful for diagnostic and prognostic purposes, no study has until now found a single marker that alone can discriminate between bacterial and viral meningitis or that alone predicts the outcome of bacterial meningitis. However, protein array analysis of CSF from patients with meningitis may be an interesting screening method for new diagnostic and prognostic CSF markers (Kastenbauer et al 2005).

5. PROGNOSIS AND OUTCOME

The bacterial pathogen itself is found to be an important determinant for the clinical outcome of meningitis with *S. pneumoniae* resulting in the highest mortality and morbidity among pathogens causing community-acquired meningitis in developed countries (e.g. *S. pneumoniae*: ~25%, *Neisseria meningitidis*: ~5-10% (van de Beek et al 2004)). Among cases with pneumococcal meningitis, we also showed that serotype-related differences in mortality existed (**Figure 3**, (Østergaard et al 2004b)), whereas the outcome did not rely on the susceptibility of the pneumococcal isolate (Fiore et al 2000; Kellner et al 2002; Østergaard et al 2005). Future testing of clinical pneumococcal CSF isolates for other virulence factors in relation to outcome may determine important targets for new protein based pneumococcal vaccines.

Table 1. Selected studies of CSF inflammatory mediators.

CSF analysis	Authors	Comments
Expression of TNF α and transforming growth factor (TGF) β -1	Ossege et al 1994	In situ hybridisation
Expression of TNF α , interferon (IFN) γ , IL-1, TGF β , endothelin-1	Rieckmann et al 1995	Semi-quantitative RT-PCR
ΤΝFα	Lopez-Cortes et al 1993	Differential value
Caspase-1	Koedel et al 2002b	
IL-1β,	Mustafa et al 1989	Prognostic value
sTNFαR	Ichiyama et al 1996	Differential and prognostic value
$TNF\alpha$ and $TGF\beta\text{-}1$	Ichiyama et al 1997	Prognostic value
IFNγ	Ohga et al 1994	
IL-6	Chavanet et al 1992	
IL-8	Østergaard et al 1996	Differential value
GRO α , monocyte chemotactic protein-1, macrophage inflammatory	Sprenger et al 1996	
protein (MIP)-1 α , and RANTES		
IL-10	Frei et al 1993	Differential value
IL-12, IFNγ	Kornelisse et al 1997	
IL-2, SIL-2K	Larsen and Bjerager 1990	
IL-18 and IFNγ	Fassbender et al 1999	
Macrophage migration inhibitory factor	Østergaard et al 2002a	
scD14	Nockner et al 1999	
st-selectin and selarity melocyle (ICAM) 1	Fassberider et al 1997	
	Lewczuk et al 1998	
CP	Stearman and Southgate 1994	
Matrix motallegrateinases (MMD)	Lopport et al 2000	Prognastic value
Nitric oxido	Korpolice et al 1996	Prognostic value
S fragtalking	Kortonbauer et al 2002	
	Østorgaard of al 2003	Prognostic value
Γ.L-40	Østergaard et al 2002b	Prognostic value
Follictin	Michol et al 2000	
a-melanocyte-ctimulating hormone	Ichivama et al 2000	Prognostic value
Glutamate	Spranger et al 1996	Prognostic value
Nitrotyrosine	Kastenhauer et al 2002a	Prognostic value
	Rastenbauer et al 2002a	



Serotype 9V (n=59)

Figure 3. Kaplan-Meier survival curve of pneumococcal meningitis according to serotypes. Significant difference between groups (Log rank test: P=0.0047). Patients infected with serotype 3 and 9V had significantly higher mortality rate than patients infected with serotype 1 (Log rank test P=0.0065 and P=0.0006, respectively).

Two out of every three deaths from pneumococcal meningitis occurred within the first week of hospitalisation, but death related to the disease was observed up to 3 months after admission (Østergaard et al 2004b; Østergaard et al 2005; Weisfelt et al 2006). Therefore, our results may suggest a longer study period when studying pneumococcal meningitis than a mortality at 14 days as the endpoint, as previously suggested for studying community-acquired bacterial meningitis (McMillan et al 2001). The causes of death from pneumococcal meningitis were due to both neurological complications (e.g. increases intracranial pressure, brain oedema, cerebral incarceration in ~50%) and to systemic complications (e.g. septic shock, stress haemorrhagic ulcers, multiorgan failure in ~50%), and neurological sequelae such as hearing loss, mental retardation, limp paralysis occurred in up to half of survivors, (Bohr et al 1985; Kastenbauer and Pfister 2003; Østergaard et al 2005; Weisfelt et al 2006).

Several studies have tried to identify risk factors associated with fatal outcome and the development of sequelae from pneumococcal meningitis, but because most studies have been relatively small in size and included different or selected study populations (e.g. adults vs. children or patients admitted to intensive care units (Auburtin et al 2002), respectively), results have differed between studies. However, the case fatality rate of pneumococcal meningitis has consistently been shown to be twice as high in developing countries (Baird et al 1976; Goetghebuer et al 2000; Gordon et al 2000) as in industrialised countries (Østergaard et al 2005), to be higher in adults (Kastenbauer and Pfister 2003; Weisfelt et al 2006) than in children (Casado-Flores et al 2005; Fiore et al 2000; Kornelisse et al 1995; Laxer and Marks 1977), and to be associated with advanced age among adult cases (Bohr et al 1985; Kastenbauer and Pfister 2003; Østergaard et al 2005; Weisfelt et al 2006) (Figure 2). Less consistent findings include an increased risk of fatal outcome in cases, who had an accompanying underlying disease, who had an altered mental status on admission, or who developed complications during hospitalisation (e.g. seizures, need for assisted ventilation), respectively (Bohr et al 1985; Henneberger et al 1983; Kornelisse et al 1995; Østergaard et al 2005; Weisfelt et al 2006; Weiss et al 1967). Moreover, presence of bacteraemia or pneumonia has been associated with increased mortality (Bohr et al 1985; Kastenbauer and Pfister 2003; Laxer and Marks 1977), whereas we showed that presence of an otogenic focus conversely was associated with a lower case fatality rate (Figure 4, (Østergaard et al 2005)). Furthermore, meningitis



Figure 4. Kaplan-Meier survival curve of pneumococcal meningitis according to the focus of the infection. Significant difference between groups (Log rank test: P=0.0005). Otogenic focus vs. pneumonic focus, sinusitic focus, other foci, and no primary infection focus: P=0.0002, 0.008, <0.0001, and 0.03, respectively. Other foci vs. no primary infection focus: P=0.01

patients receiving antibiotics prior to admission were found to have a lower mortality than patients, who did not receive antibiotics (Bonsu and Harper 2001; The Research Committee of the BSSI 1995), and a delay in diagnosis and start of antibiotic therapy (e.g. due to CT-scan before diagnostic lumbar puncture) during hospitalisation was associated with worsened outcome (Aronin et al 1998; Østergaard et al 2005), emphasising the importance of prompt initiation of antibiotic therapy.

6. PATHOLOGY

Percent survival

Autopsy studies performed in the pre-antibiotic era (Southard and Keene 1906) and from patients receiving antibiotic therapy (Cairns and Russell 1946; Quade and Kristensen 1962) have shown that pneumococcal meningitis is characterised by inflammation within the subarachnoidal space and along the cerebral vasculature predominantly of neutrophil origin and various degrees of histopathological alterations within the brain parenchyma. The inflammatory reaction within the subarahnoidal space appears as purulent exudates on the surface of the brain, and the inflammatory reaction around the vessels appears as artheritis and phlebitis with thrombosis occasionally observed within the vascular lumen. In addition, abscess formation may be found around inflamed vessels affecting the surrounding brain parenchyma. Within the brain parenchyma, brain oedema and signs of cerebral incarceration due to increased intracranial pressure may be found as well as focal cortical necrosis and ischemic lesions. In hippocampus neural apoptosis is observed in the dentate gyrus (Nau et al 1999a). Only a limited number of brain autopsies is available, which represents a selected material from only fatal cases, and which may be of poor technical quality due to the degenerative processes taking place after death, until the brain is preserved. Therefore, animal models are essential for studying the pathogenesis and pathophysiology of bacterial meningitis and in elucidating optimal antibiotic and adjunctive therapies.

7. ANIMAL MODELS

Optimally, the animal model should resemble the natural course of human pneumococcal meningitis (e.g. route of infection, histopathological alteration, systemic complications), but the experimental design may depend on the scientific issue studied and on the end-point monitored. In all experimental animal models, CNS bacterial invasion or inoculation of pneumococci directly into the CNS causes an accumulation of leukocytes within the subarachnoidal space, increased blood/brain barrier permeability, development of brain oedema and increased intracranial pressure. In contrast, the causes of death and degree of brain damage vary according to the model (O'Reilly et al 2005).

The mouse meningitis model has been used to study the pathogenesis of pneumococcal meningitis, because it has been possible to induce meningitis after intranasal inoculation either with or without the use of hyaluronidase as facilitating agent (Orihuela et al 2003; Zwijnenburg et al 2001). Also, two alternative routes of pneumococcal inoculation (intracisternal (Echchannaoui et al 2002; Koedel et al 2003) and intracerebral (Gerber et al 2001)) have been used. An increasing number of studies using gene-modulated knockout mice have recently been performed and have been very useful in the study of host immune reactions (Paul et al 2003a). Cortical brain damage (Klein et al 2006) and neural apoptosis in hippocampus (Mitchell et al 2004) is observed in this model.

The rabbit meningitis model has been used extensively in studying CSF dynamics (CSF penetration of antibiotic, CSF bacterial growth – or kill rate, CSF components of the meningeal inflammatory response) during the course of pneumococcal meningitis, because sequential tapping is possible (Dacey and Sande 1974). Another advantage of this model is that the immune response phylogenetically is closer to man than for other rodents (Graur et al 1996), and IL-8 is present in rabbits (Harada et al 1993) contrary to in mice and rats. We adjusted this model, so that it was possible to study the meningeal inflammatory response in the early phase of pneumococcal

meningitis before and during the start of CSF pleocytosis to identify important regulatory mediators (Østergaard 2000). However, the rabbit model has limitations in studying mortality and brain damage, because a rapid secondary bacteraemia develops, and death primarily occurs due to systemic complications (e.g. septic shock, ARDS) (Stewart 1927). In addition, no brain damage has been found in the cortex (**Figure 5**), whereas neural apoptosis in hippocampus has been detected (Braun et al 1999; Zysk et al 1996). On the other hand, the rabbit model has been useful in the study of hearing loss due to meningitis (Bhatt et al 1993).

The rat meningitis model. The infant rat meningitis model has consistently shown histopathological alterations close to human pneumococcal meningitis with both cortical and hippocampal involvement (Leib et al 2000; Leib et al 2003). Less consistent results have been obtained in the adult rat model, where we and others have established a model with histological alteration that include significant cortical involvement (e.g. focal cerebral abscess formation and cortical necrosis, (Figure 5)) (Brandt et al 2004; Tauber et al 1992). Others find less significant brain damage and have monitored intracranial pressure, brain oedema and blood/brain barrier permeability as endpoint parameters (Koedel et al 1995). Discrepancies could be due to difference in virulence of the pneumococci used, since this has been crucial for the development of brain damage in our model (Brandt et al 2004). Indeed, distinct differences in degree and pattern of brain damage was observed with the use of different serotypes in the adult rat model (serotype 1: vasculitis and cortical



Figure 5. Brain damage in rabbits and rats due to 5. pneumoniae meningitis. No brain damage was observed in rabbits (upper left), whereas rats had various degree and pattern of cortical involvement, i.e. ischaemia/necrosis (upper right), abscess formation, vasculitis and haemorrhagia (lower feft) as well as subcortical abscess formation (lower right).

Figure 6. Cochlea damage due to experimental *S. pneumoniae* meningitis in rats. Section of the spiral ganglion from the basal turn of the cochlea. Comparison between a control animal (A) and a G-CSF pre-treated animal (B), which displays a nearly complete loss of spiral ganglion neurons. PAS-alcian blue staining (Brandt et al 2006a).

haemorrhagia, serotype 3: cortical necrosis and abscess formation, and serotype 9V: subcortical (callosal) abscess formation (Østergaard et al 2004b)). Significant hippocampal involvement has not been observed in the adult rat model, which could indicate that adult neurons are less susceptible to neuroexcitatory stress than neonatal neurons. The adult rat model has also been valuable in the study of hearing loss (Brandt et al 2006a; Klein et al 2003) (Figure 6).

8. PATHOGENESIS

Sternberg and Pasteur discovered the S. pneumoniae simultaneously in 1881 (for a review: (Austrian 1981a)). Important pathogenic structures of the pneumococcus are the polysaccharide capsule (variation in its antigenic structure allows to distinguish in more than 90 different serotypes according to the Danish nomenclature) and the cell wall. A thorough morphological decription of the various structural components of the pneumococcus has previously been presented in a Danish Doctoral thesis (Sørensen 1995). One important characteristic of the pneumococcus is its ability to adapt to the environment and up- and down regulate several surface structures that help the pneumococcus to survive and even take advantage of host reactions (Gillespie and Balakrishnan 2000). Such a variation between different phases can be visualised phenotypically by its morphological colony appearance on agar plates, and transparent and opaque colonies were found to express large amount of cell wall (e.g. choline binding proteins) and capsule (polysaccharides), respectively (Cundell et al 1995b).

No clinical studies have until now demonstrated the exact route and mechanism for developing pneumococcal meningitis, but theoretically it may either be through direct invasion from a primary infection focus close to the meninges (e.g. ear, sinus, dura disruption) or through haematogenous spread from a distant infection focus (e.g. lung, nasopharynx colonisation). To further address this issue we studied the presence of bacteraemia according to the focus of the infection and found that bacteraemia was observed in $\sim 67\%$ of cases with pneumococcal meningitis with no significant difference between cases with a close or a distant primary infection focus (Østergaard et al 2005), indicating that pneumococcal meningitis predominantly may be introduced haematogenously. However, secondary bacteraemia is found to occur within ~4-8 hours after intracisternal inoculation of animals with pneumococci (Østergaard 2000). Therefore, studies with quantitative cultures of CSF and blood (La Scolea and Dryja 1984) are still needed in cases with pneumococcal meningitis due to different foci to yield additional information about pneumococcal CNS invasion.

It is well-known that the pneumococcal capsule is an important virulence factor, since all invasive pneumococcal isolates are encapsulated to survive the innate immune response within the systemic compartment (Austrian 1981b). Moreover, some serotypes has from unidentified reasons - more frequently been isolated from patients with meningitis (e.g. 12F, 6B) than from patients with bacteraemia (e.g. 1 and 14 (Hausdorff et al 2000; Konradsen and Kaltoft 2002)). Sparse information about other pneumococcal virulence factors has yet been generated from clinical isolates, among these, strains isolated from patients with pneumococcal infection (including meningitis) were more frequently producing hyaluronidase than carrier strains (Kostyukova et al 1995). However, significant new knowledge about pneumococcal virulence factors has been obtained experimentally. Signature-tagged mutagenesis (Polissi et al 1998) and micro-array analysis (Orihuela et al 2004b) have provided useful screening methods for potential pneumococcal virulence factors that may lead to the design of genetically modulated pneumococcal mutants for further testing in in vitro models and in animal models. Moreover, advances in bioluminescence imaging has provided a useful non-invasive method for studying the progression of pneumococcal meningitis in vivo (Kadurugamuwa et al 2005) and in a mice model it was found that serotype-related differences in the ability to

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cause meningitis exist after intranasal inoculation of various serotypes (Orihuela et al 2003). Furthermore, a recent study has thoroughly described the role of various virulence factors in the integrated processes taking place from nasopharyngeal colonization eventually leading to CNS invasion (Orihuela et al 2004a):

Colonization and epithelial transmigration. Pneumococci produce IgA proteases able to degrade mucosal IgA, which may protect them from the mucosal immune system (Poulsen et al 1998). Pneumococci produce neuraminidases that enhance epithelial adherence (Tong et al 2000). Pneumococci upregulate cholin-binding proteins (e.g. CbpA) that bind to epithelial polymeric immunoglobulin receptor, which is important for the adherence and the migration of pneumococci through epithelial cells (Cundell et al 1995a; Zhang et al 2000). In experimental meningitis, neuraminidase, pyruvate oxidase and CbpA all contributed to nasopharyngal colonisation, whereas pneumolysin, pyruvate oxidase, and autolysin contributed to further systemic invasion (Orihuela et al 2004a). Also, hyaluronidase facilitated systemic invasion after intranasal inoculation of pneumococci, probably by degrading the basement membrane (Zwijnenburg et al 2001).

Survival in the systemic circulation. For survival in the bloodstream, pneumococci upregulate the expression of capsular polysaccharide (Kim and Weiser 1998) and release pneumolysin, resulting in an attenuation of the innate immune response (Gillespie and Balakrishnan 2000). In experimental meningitis, pneumolysin and autolysin contributed to an increased degree of bacteraemia (Orihuela et al 2004a).

CNS invasion. Upregulation of CbpA facilitates binding to the endothelial platelet activating factor receptor and subsequent transmigration of pneumococci through the endothelial cell into the CNS as shown in vitro (Ring et al 1998) and in experimental meningitis (Orihuela et al 2004a). Once entered the CSF, an exponential growth of pneumococci may occur as shown in experimental meningitis models. However, the CSF is a poor growth media for pneumococci, as shown in **Figure 7**.

Virulence factors and outcome. A direct causal role of the pneumococcal toxins (e.g. pneumolysin and H2O2, pneumococcal adherence and virulence factor A) for mortality and for the development of hippocampal neural apoptosis has been documented in experimental meningitis using genetically modulated mutants of pneumococci, whereas neuraminidase- and hyaluronidase deficient strains did not influence the outcome (Braun et al 2002; Pracht et al 2005; Wellmer et al 2002). As shown in human pneumococcal meningitis, an association between CSF concentrations of lipoteichoic/teichoic acid and a poor outcome has also been documented experimentally (Gerber et al 2003; Nau et al 1999b).

9. PATHOPHYSIOLOGY

The immune reaction in pneumococcal meningitis against the invading pathogen includes both local - and systemic host inflammatory reactions and involves both innate and adaptive immune responses. An important characteristic of the immune response within the CNS that clearly differentiates it from the systemic immune response is an impaired opsonic - and phagocytic activity (Simberkoff et al 1980) and the incapability of controlling the infection within the CNS compartment, which uniformly leads to the death of the patients. Particularly, the adaptive immune response is impaired within the CNS compartment and active immunization with heat-killed pneumococci intracisternally as well as intravenously did not induce a significant intrathecal antibody production able to attenuate CSF bacterial growth contrary to the significant systemic antibody production after immunization capable of controling the systemic infection (Østergaard et al 2006b; Stewart 1927). Also, the innate immune response is impaired in the CNS compartment (e.g. impaired complement activity (Crosson, Jr. et al 1976; Zwahlen et al 1982)). Therefore, intravenously and/or intrathecally administered serotype-specific antibodies experimentally



Figure 7. CSF bacterial growth and bactericidal activity of penicillin. A. Time-kill studies of *S. pneumoniae* in broth and in human CSF. B. Bacterial concentrations in CSF from rabbits infected with *S. pneumoniae*. Arrows indicate dosing of penicillin (0.16 MIE/kg as 1. dose and 0.08 MIE/kg as 2. dose).

used in the pre-antibiotic era did not lead to the survival of patients with pneumococcal meningitis (Kolmer 1929), which was in contrast to the better outcome of systemic pneumococcal infection with serum therapy (Avery et al 1917). But besides impaired infection control by the CNS host response, which can be compensated with antibiotic therapy, the host response may cause harmful effects, since injections of heat-killed pneumococci into cisterna magna induced pathophysiological CNS alterations resulting in the death of rabbits (Tuomanen et al 1989).

A complex cascade of events takes place with the activation of the innate immune system that includes recruitment of leukocytes from the systemic compartment and release within the CNS of inflammatory mediators (e.g. cytokines, chemokines, reactive oxygen radicals, excitatory amino acids and proteolytic enzymes). Experimental studies have investigated modulation of the various steps in the inflammatory cascade of pneumococcal meningitis to describe a functional role. These studies include pharmacological intervention and recently a number of studies using knock out mice have been performed.

Activation. Intracisternal inoculation with live or heat-killed pneumococci or various pneumococcal components are capable of inducing a meningeal inflammatory response in animals that mimic the meningeal alterations observed in patients with pneumococcal meningitis (Tuomanen et al 1985a). Various experimental procedures influenced the extent and the pattern of meningeal inflammatory response (e.g. CSF pleocytosis, cytokine kinetics) during experimental pneumococcal meningitis (e.g. inoculum size, the use of heat-killed or living bacteria, serotypes, antibiotic therapy) (Østergaard 2000). For example, inoculation of high inoculum sizes of living serotype 3 (~ 10^8 CFU = ~ 10^7 CFU/mL CSF) induced a proinflammatory cytokine response with TNF α and IL-1 β preceding the pleocytosis, which was not observed during CSF bacterial growth after inoculation with a low inoculum size (Figure 8). Also, intracisternal injection of pneumococcal cell wall material (> $0.2 \mu g$) and serotype 3 capsule (>200µg) as well as LPS from Gram-negative bacteria induced a proinflammatory cytokine response preceding the pleocytosis ((Tuomanen et al 1985b), Østergaard, C., unpublished data). Moreover, inoculation of heat-killed as compared to living pneumococci resulted in an enhanced proinflammatory cytokine response, however with a significantly attenuated pleocytosis (Figure 8). Various components of the pneumococcal cell wall (teichoic acid and peptidoglycan) have been identified as main inducers of the meningeal inflammatory response during experimental meningitis (Tuomanen et al 1985a).

The molecular explanation for the recognition of pneumococci and the downstream activation of the innate immune response has been studied extensively in vitro and recently in experimental meningitis using knockout mice. LPS binding protein (LBP), an acute phase protein that binds to pneumococcal peptidoglycans causing an activation of the innate immune system through Toll-like-receptor (TLR)-2, played a biological role in pneumococcal meningitis, since LBP was upregulated during pneumococcal meningitis, and LBP-deficient mice had an attenuated pleocytosis that could be restored after intrathecal injection of recombinant LBP as compared to wild type mice (Weber et al 2003). TLR's were upregulated in the CNS during experimental pneumococcal meningitis (TLR-2, 4 and 9 mRNA) (Bottcher et al 2003b), however TLR-2 deficient mice showed no difference in pleocytosis compared to wild type mice (Echchannaoui et al 2002; Koedel et al 2003) suggesting that other TLR's than TLR-2 are involved in activation of the innate immune response in pneumococcal meningitis. Indeed, intracisternal inoculation of bacterial DNA induced pleocytosis primarily of monocytic origin (Deng et al 2001) and most likely through TLR-9 (Hemmi et al 2000). The intracellular transmission of the activation signal from membrane-bound TLR to the nuclear transcriptional factor is mediated through myeloid differentiation factor 88 (MyD88), and in pneumococcal meningitis, MyD88-deficient mice had an attenuated pleocytosis and cytokine response (Koedel et al 2004). The intracellular transcription factor- κB (NF κB) is the final step in the activation by the invading pathogen resulting in the subsequent production of inflammatory mediators and upregulation of adhesion molecules promoting leukocyte recruitment into the CNS, and NFkB was upregulated in experimental pneumococcal meningitis, and inhibition of NFkB attenuated both pleocytosis and CSF cytokine response (Koedel et al 2000).

Adhesion and migration. With activation of NFκB, selectin adhesion molecules are upregulated on the luminal surface of the endothelial cells, which perform a weak and incomplete binding to selectins on the leukocyte resulting in leukocyte rolling along the cerebral vasculature. In experimental pneumococcal meningitis, therapy with the selectin-blocker fucoidin inhibited leukocyte rolling along endothelial cells (Granert et al 1994) and attenuated the pleocytosis (Angstwurm et al 1995; Granert et al 1998; Granert et al 1999; Østergaard et al 2000b). Also, P- and E-selectin deficient mice had decreased pleocytosis after cykine-induced meningitis (Tang et al 1996), and therapy with peptides derived from pertusis toxin, causing a competitive inhibition of the binding to selectins, reduced pleocytosis in experimental pneumococcal meningitis (Rozdzinski et al 1993; Sandros et al 1994).

The next step in the adhesion process is the strong binding between integrins on the leukocytes and immunoglobulins on the endothelial cells resulting in firm attachment. Therapy with mono-



Figure 8. Inoculum size and meningeal inflammatory response during experimental pneumococcal meningitis. CSF kinetics in rabbits infected with S. pneumoniae (10⁵ CFU (n=7), 10⁶ CFU (n=7), 10⁸ CFU (n=5), 10⁸ CFU heat-killed (n=10)).

clonal antibodies to CD18 (b2 integrin) (Tuomanen et al 1989; Zysk et al 1996) or therapy with peptides derived from filamentous hemagglutinin of Bordatella pertusis, causing an competitive inhibition of binding to integrins, attenuated pleocytosis in experimental pneumococcal meningitis (Rozdzinski et al 1995a; Rozdzinski et al 1995b). Also, therapy with monoclonal antibodies to the endothelial immunoglobulin, CD54 in experimental pneumococcal meningitis (Weber et al 1995) or to junctional adhesion molecule in cytokineinduced meningitis attenuated the pleocytosis (DelMaschio et al 1999).

The transmigration of leukocytes through the endothelial cells into the CNS compartment is induced by local chemotactic gradients (e.g. chemokines as discussed below) and by a release of proteolytic enzymes from the leukocyte (e.g. gelatinase, elastase, uPAR) breaking down the basement membrane on the apical side of the endothelial cell. Intracerebral injection of MMP resulted in blood/brain barrier disruption (Lukes et al 1999), and therapy with MMP inhibitors reduced blood/brain barrier disruption, but not pleocytosis in experimental meningitis (Paul et al 1998), whereas MMP-deficient mice had no significant difference in pleocytosis and blood/brain barrier permeability as compared to wild type mice during pneumococcal meningitis (Bottcher et al 2003a). Also, intracisternal injection of elastase increased blood/brain barrier permeability, but only marginally pleocytosis (Temesvári et al 1995). In contrast, uPAR deficient mice had an attenuated pleocytosis, but no alterations in blood/brain barrier permeability in experimental pneumococcal meningitis, suggesting a chemotactic role of uPAR (Paul et al 2005).

Cytokines and chemokines. Cytokines and chemokines are produced by local cells (e.g. endothelial cells, astrocytes, microglial cells) and by blood-derived leukocytes, and they play a key role in endothelial activation and leukocyte recruitment and function (Tauber and Moser 1999). A functional role of cytokines and chemokines as well as the cellular origin of cytokine production has been investigated in experimental pneumococcal meningitis (Zwijnenburg et al 2006). However, conflicting results have been obtained, which may be due to species differences and/or to differences in methods used.

In the rabbit model, expression of TNFα and IL-1β was primarily observed in mononuclear cells within the cellular infiltrate (Bitsch et al 1997), whereas preliminary results showed that IL-8 was predominantly detected along the cerebral vasculature (C. Østergaard and D. Hougaard, unpublished data). Pleocytosis has been induced with intracisternal injection of rabbit TNFα and IL-1β (Ramilo et al 1990b), human MIP-1 and 2 (Saukkonen et al 1990), whereas rabbit or human IL-8 did not induce pleocytosis (Dumont et al 2000; Østergaard et al 2000a). Also, therapy with monoclonal antibodies against TNFα (i.c.), IL-1β (i.c.) , IL-8 (i.v., but not i.c.) (Dumont et al 2000; Østergaard et al 2000a; Saukkonen et al 1990) or the inhibition of IL-1β by caspase inhibition (Braun et al 1999) attenuated CSF pleocytosis in experimental pneumococcal meningitis. In contrast, therapy with IL-10 did not attenuate pleocytosis but did reduce CSF TNFα levels in experimental meningitis (Paris et al 1997).

In the rat model, various cytokines are upregulated during pneumococcal meningitis (Diab et al 1997). Pleocytosis was induced by injection of human TNF α and IL-1 β intracisternally (Quagliarello et al 1991) and intraspinally, but not intracerebrally (Schnell et al 1999), and in experimental pneumococcal meningitis pleocytosis was attenuated by IL-10 (i.v. but not i.c.) (Koedel et al 1996) and by caspase inhibition (Koedel et al 2002b).

In the mouse model, intracerebral injection of TNF α , IL-1, IL-8, PAF (Andersson et al 1992) and intracisternal injection of MIP-2

and KC (Zwijnenburg et al 2003a) resulted in pleocytosis. However, mice deficient of TNF α or its receptors (Gerber et al 2004; Wellmer et al 2001), mice deficient of IL-1R type 1 (Zwijnenburg et al 2003c), mice deficient of IL-10 (Zwijnenburg et al 2003d) and mice deficient of IL-18 (Zwijnenburg et al 2003b), respectively, had no attenuation in CSF pleocytosis in experimental pneumococcal meningitis. In contrast Caspase-1 deficient mice had attenuated pleocytosis (Koedel et al 2002b), whereas IL-6 deficient mice had enhanced CSF pleocytosis (Paul et al 2003b).

Influence of the systemic infection/inflammation on the meningeal inflammatory response. The CSF pleocytosis is influenced by events taking place within the systemic compartment: There was significant correlation between blood WBC and CSF WBC in patient with pneumococcal meningitis (Østergaard et al 2006b), and in experimental pneumococcal meningitis an attenuated CSF pleocytosis was observed with the induction of leukopenia (Ernst et al 1983) or after depletion of mononuclear cells in the systemic compartment (Zysk et al 1997b), but not after depletion of CNS macrophages (Trostdorf et al 1999). Moreover, an earlier onset of bacteraemia caused an attenuated pleocytosis most likely due to a decrease in number of peripheral leukocytes (Østergaard et al 2006b), and also G-CSF pretreatment resulted in an attenuated pleocytosis most likely due to decreased chemotactic ability of leukocytes (Østergaard et al 1999).

In meningitis patients, high TNF α and IL-1 β levels were detected in the CSF with low or not detectable corresponding levels in the blood, whereas no pleocytosis was observed during bacteraemia with significantly lower CSF cytokine levels as compared to blood levels (Waage et al 1989). In further support that cytokines are released locally in the CSF during meningitis, a net efflux of cytokines from the brain to the blood was observed in patients with pneumococcal meningitis (Møller et al 2005) contrary to no observed efflux of cytokines after intravenously injection of LPS (Møller et al 2002). In experimental pneumococcal meningitis, attenuation of the CSF pleocytosis, however, resulted in decreased CSF levels of cytokines predominantly produced by blood-derived cells (e.g. TNFa (Bitsch et al 1997; O'Reilly et al 2007), IL-1 (Østergaard et al 2000b; Zysk et al 1997b), whereas cytokines produced by local cells (e.g. IL-8) responsible for the chemotactic signal were augmented (Østergaard et al 2000b; Østergaard et al 2006b).

Leukocyte activation. With the development of pleocytosis several pathophysiological events occur that include both direct cytotoxic alterations and vascular alterations. The exact causal and temporal role of these events are not completely elucidated, but when entering the CNS, the leukocyte is able to release several activation products in the defence against invading microorganisms (Borregaard 1996). These include reactive oxygen species and nitric oxide, which were markedly elevated during experimental pneumococcal meningitis (Koedel and Pfister 1999). However, such products are not only toxic for the microorganisms but also for the host cells, and besides the effects on brain tissue, nitric oxide and reactive oxygen radicals also affect endothelial cells and cause changes in the tonus of the cerebral vasculature (vasodilatation and vasoconstriction, respectively), as will be described below.

Brain oedema and increased intracranial pressure. With the disruption of the blood/brain barrier that normally forms a tight barrier between the systemic compartment and the CNS, an increased permeability (elevated CSF protein levels) may lead to brain oedema (Quagliarello et al 1986). Blocking of leukocytes entry into the CSF reduced CSF protein levels (Østergaard et al 2000b; Tuomanen et al 1989), whereas a decrease in peripheral WBC was not associated with decreased blood/brain barrier permeability in experimental pneumococcal meningitis (Ernst et al 1983; Østergaard et al 2006b; Tauber et al 1988). Also, an increased outflow resistance most likely caused by the meningeal inflammation may participate in the development of brain oedema (Scheld et al 1980). Recent MR-findings showed that during experimental pneumococcal meningitis, vasogenic (extacellular) oedema is the earliest event observed followed by a shift to cytotoxic (intracellular) oedema (Brandt et al 2006b). Because the rigidity of the cranial cavity limits the expansion of the brain volume, brain oedema may result in an increased intracerebral pressure that may lead to fatal brain herniation and/or a global decrease in cerebral perfusion/blood flow. Osmotic therapy (glycerol, manitol, diuretics) may be tried to reduce ICP, however, the documentation of treatment efficacy is still lacking (van de et al 2006). Also, continuous measurements of ICP after placement of a ventricular shunt with active intervention against elevated ICP levels may be a promising future treatment option (Grande et al 2002; Lindvall et al 2004) in bacterial meningitis.

Cerebral blood flow. After an increase in cerebral blood flow initially (Pfister et al 1990b), further progress in the disease resulted in a global decrease in cerebral blood flow as observed in experimental pneumococcal meningitis (Tauber et al 1991) and in meningitis patients (Paulson et al 1974). A loss of cerebral autoregulation was found in patients with bacterial meningitis (Møller et al 2001b) and in experimental pneumococcal meningitis (Tureen et al 1990). Consequently, the induction of systemic hypotension was recently shown to result in a decrease in global cerebral blood flow during experimental pneumococcal meningitis, whereas an augmentation of the mean arterial blood pressure by norepinephrine increased cerebral blood flow (Pedersen et al 2007). Interestingly, hyperventilation partially restored normal autoregulation in experimental pneumococcal meningitis (Pedersen et al 2007) and in meningitis patients (Møller et al 2000b). Also, hydration status influenced global cerebral perfusion in experimental pneumococcal meningitis (Tureen et al 1992).

Focal cerebral perfusion abnormalities were found in patients with bacterial meningitis (Förderreuther et al 1992; Møller et al 2000a), and the focal nature of brain damage (e.g. wedge-shape necrosis/ischemia around occluded vessels) found in pneumococcal meningitis (Brandt et al 2004) suggest that several mediators locally released during the inflammatory process may influence local cerebral perfusion. Indeed, inhibition of mediators causing vasoconstriction such as superoxide radicals (Auer et al 2000; Koedel and Pfister 1997) and endothelins (Koedel et al 1998; Pfister et al 2000) increased cerebral blood flow and attenuated development of brain damage in experimental pneumococcal meningitis. Inhibition of mediators causing vasodilatation such as nitric oxide (NO) and endothelial NO synthase (NOS) decreased blood flow and increased ischemic brain damage (Koedel et al 1995; Koedel et al 2001), whereas deficiency of inducible NOS was beneficial in experimental pneumococcal meningitis (Winkler et al 2001).

Neurotoxicity. As documented in stroke models, ischemiacally induced elevation in CNS concentrations of neuroexitatory amino acids has been detected (Guerra-Romero et al 1993) and has been shown to contribute to development of brain damage in experimental meningitis (Leib et al 1996b).

10. THERAPY

Antibiotics. The most important step in the treatment of bacterial meningitis is the prompt initiation of antibiotic therapy. Randomised clinical meningitis trials evaluating different antibiotic treatment regimes have been performed (Saez-Llorens et al 2002; Schaad et al 1990), but a Cochrane review showed no difference in clinical outcome between the use of broad-spectrum and narrowspectrum antibiotics (Prasad et al 2004). Therefore, the choice of antibiotic therapy for treatment of bacterial meningitis depends primarily on local susceptibility patterns for meningeal pathogens, the age of the patient, and on considerations of CSF pharmacokinetic and pharmacodynamic properties of antibiotics. Such pk/pd data have predominantly been obtained from experimental studies using the rabbit meningitis model. Empiric therapy with a third generation cephalosporin in combination with penicillin will cover most meningeal pathogens in Denmark (Meyer et al 2004), whereas in countries with high penicillin - and cephalosporin resistance, recommended therapy includes the addition of vancomycin and/or rifampicin (Tunkel et al 2004), but also therapy with newer fluoroquinolones (e.g. moxifloxacin) should be considered (Saez-Llorens et al 2002).

CSF penetration. The blood/brain barrier forms a tight membrane that limits free passage of antibiotics into the CNS. The penetration of antibiotics across the blood/brain barrier into the CNS was found to be facilitated by high lipid solubility (lipophilic drugs: fluoroquinolones, rifampicin, metronidazole, hydrophilic drugs: b-lactams) and by low molecular weight (i.e. high: vancomycin, low: fluoroquinolones) of the drug (Nau et al 1994), whereas the role of protein binding has not been fully elucidated. Inhibition of an active efflux pump by probenecid increased the CSF penetration of penicillin (Dacey and Sande 1974). However, the most important factor for the CSF penetration 10-fold (**Figure 9**) (O'Reilly et al 2005; Østergaard et al 1998; Østergaard et al 2003). The CSF penetration of various antibiotics obtained in the rabbit meningitis model is shown in **Figure 10**.

CSF bacterial killing. CSF bactericidal activity of antibiotics is compromised because of poor bacterial growth rate in CSF (Figure 7). Therefore, higher antibiotic concentrations are required to obtain maximal bacterial killing within the CSF than in the systemic compartment, also for drugs normally showing minimal concentration-dependent killing (e.g. β -lactams: ~10-100 × the MBC vs. ~4 × the MBC, respectively (Tauber et al 1984)). It is possible to sterilise the CSF within ~10-12 hours after start of antibiotic therapy for most antibiotics in the therapy of pneumococcal meningitis (Kanegaye et al 2001), however, regrowth may occur with premature withdrawal of antibiotic therapy (Østergaard et al 1998). In addition, clinical treatment failures have been observed with vancomycin, which has been explained by a lower CSF penetration of the drug with the resolvement of meningitis and the restoration of the blood/brain barrier integrity (Viladrich et al 1991).

Pd properties. Several aspects influence, why pd properties in bacterial meningitis differ from "normal" pk/pd: 1) The fluctuation in antibiotic concentration is less pronounced in the CSF than in the blood due to a \sim 3-4 times longer elimination half life in CSF than in serum, resulting in almost steady state pk/pd. 2) Complete sterilization of the infection is needed, because the immune system within the CNS cannot help clearing the bacterial infection, as discussed in detail above.

For concentration-independent antibiotics, $T_{>MBC}$ was the pd parameter correlating best with CSF bacterial killing during ceftriaxone therapy (Lutsar et al 1997). However, treatment with ceftriaxone and other β -lactams still caused additional CSF bacterial killing at CSF peak concentrations 10-100 × MBC (Tauber et al 1984). In contrast to this, therapy with vancomycin caused no additional killing at CSF concentrations 4 × MBC (Ahmed et al 1999).

For concentration-dependent antibiotics (e.g. fluoroquinolones) additional killing occurred at concentrations at least $40 \times MBC$. AUC/MBC was the pd parameter correlating best with CSF bacterial killing during fluoroquinolone therapy, but regrowth occurred when CSF concentrations fell below the MBC (Lutsar et al 1998; McCoig et al 1999; Østergaard et al 1998).

Combination therapy. Several studies have investigated various combinations of antibiotics. In general, combination of bacterio-static and bactericidal antibiotics resulted in antagonism (Øster-gaard et al 2003), whereas combination of bactericidal antibiotics caused synergism in some studies, but indifference in others (Cot-tagnoud and Tauber 2004). The clinical importance of not combining bacteriostatic and bactericidal antibiotics was documented half a century ago, where therapy with penicillin in combination with aureomycin resulted in a higher mortality than therapy with penicillin alone (Lepper and Dowling 1951). However, recent experimental results showed that such antagonism could be compensated with the use of higher β -lactam doses (Meli et al 2006).

CSF bacterial killing and mortality. Antibiotic induced CSF bacter-

ial killing may cause an increased meningeal inflammatory response (Friedland et al 1995; Tuomanen et al 1987), and the injection of heat-killed bacteria into cisterna magna of rabbits not only induced



Figure 9. CSF antibiotic penetration through inflamed and non-inflamed meninges in rabbits. The CSF penetation (AUC_{CSF}/AUC_{blood}) of ceftriaxone, vancomycin, and moxifloxacin was 15%, 13%, and 81%, respectively for infected CSF, and 1.9%, 1.1%, and 47%, respectively for uninfected CSF.



Figure 10. CSF penetration of various antibiotics obtained in the rabbit meningitis model.

a meningeal inflammatory response as discussed above, but also led to the death of the animals (Tuomanen et al 1989). However, the pattern and extent of the meningeal inflammatory response after the initiation of antibiotic therapy are not fully elucidated, and may depend on several parameters such as the time-point for the start of antibiotic therapy (Pfister et al 1994) and the ability of the pathogen to lyse with antibiotic therapy (Tuomanen et al 1988) as well as the antibiotic used. Indeed, some antibiotics induced a high release of bacterial cell wall fragments (e.g. ceftriaxone) during CSF bacterial killing, whereas others that influence the protein synthesis (e.g. rifampicin, clindamycin) did not, but on the other hand, the antibiotic induced release of lipoteichoic acid and teichoic acid within the CSF was lower with antibiotic therapy than with no treatment (Bottcher et al 2004; Spreer et al 2003; Stuertz et al 1999). Treatment with rifampicin or clindamycin in comparison with ceftriaxone reduced brain damage and/or mortality in experimental pneumococcal meningitis (Bottcher et al 2000; Bottcher et al 2004; Nau et al 1999b), whereas treatment with moxifloxacin resulted in a similar mortality as with ceftriaxone therapy (Djukic et al 2005). Therefore, future studies should investigate, whether mortality or degree of brain damage are better parameters for antibiotic efficacy than CSF bactericidal effect for the study of pk/pd in bacterial meningitis.

Adjunctive therapy with corticosteroids. A Cochrane review including 18 randomised clinical trials evaluating corticosteroids in bacterial meningitis of 1853 patients has concluded that dexamethasone has a beneficial effect on mortality in adults and on hearing loss in children (van de Beek et al 2003). Recently, a prospective, randomised European multicentre study on adults showed a beneficial effect of dexamethasone in particular on mortality in pneumococcal meningitis (de Gans and van de Beek 2002), whereas a Malawian study on children did not (Molyneux et al 2002). Including these two trials in the analysis of the efficacy of adjunctive therapy with corticosteroids (C. Østergaard, unpublished data), there was a significant beneficial effect on survival from bacterial meningitis with the use of corticosteroids (86% (1194/1386) vs. 83% (1138/1365), OR: 1.24 (95% C.I.: 1.01-1.53), P=0.043). The beneficial effect of dexamethasone on survival was dependent on the bacterial pathogen (S. pneumoniae: n=757, OR: 1.48 (1.07-2.07), P=0.017; N. meningitidis: n=570, OR: 1.33 (0.61-2.90), P=0.455; H. influenzae: n=810, OR: 1.33 (0.82-2.14), P=0.248; other pathogens: n= 100, OR: 0.79 (0.34-1.83), P=0.582; unknown pathogen: n=205, OR: 0.75 (0.33-1.69), P=0.484), on age groups (adults: n=674, OR: 1.92 (1.29-2.86), P=0.001; children: n= 2077, OR: 1.04 (0.81-1.33), P=0.776), and on socioeconomic status (industrialised countries: n=1427, OR: 1.56 (1.01-2.41), P=0.045; developing countries: n= 1235, 1.24 (0.96-1.61), P=0.098). Among pneumococcal cases, a beneficial effect was observed only among adult patients (industrialised countries: n=195, OR: 2.26 (1.20-4.29), P=0.012; developing countries: n=91, OR: 3.14 (1.31-7.51), P=0.01), whereas no significant effect was observed among children (industrialised countries: n=110, OR: 0.71 (0.04-11.7), P=0.814; developing countries: n=361, OR: 1.22 (0.78-1.89), P=0.389). Moreover, a beneficial effect of dexamethasone may be associated with the risk of dying from the infection (Figure 11), as previously documented for the effect of anti-inflammatory therapy in sepsis studies (Eichacker et al 2002). In addition, most of the beneficial effect of dexamethasone has been related to an effect on systemic complications rather than on neurological complications (van de Beek and de Gans 2004). Therefore, further meningitis studies should evaluate whether lower doses of corticosteroids as used for treatment of septic patients would be the optimal dosing regimens, as has been documented in sepsis (Annane et al 2002).

Anti-inflammatory therapy and outcome. When studying outcome of pneumococcal meningitis using animal models, several factors have to be taken into consideration. 1) The most obvious outcome parameters would be mortality or neurological sequelae (e.g. motor performance, learning deficits, hearing loss). Also, degree of brain damage has been useful due to a direct association with mortality or sequelae (Brandt et al 2004; Leib et al 2003), and because damaged brain tissue will not recover, whereas other end-point parameters such as bacterial concentrations, meningeal inflammation, intracra-



Figure 11. Adjunctive therapy with corticosteroids and mortality rate. The studies shown on the figure include studies from a Cochrane review (van de Beek et al 2003) and studies by (de Gans and van de Beek 2002) and (Molyneux et al 2002).

nial pressure, brain oedema and blood/brain barrier permeability are not always related to mortality and will resolve with the recovery from meningitis (Koedel et al 2004). 2) The animal model should optimally have a similar mortality rate as in human pneumococcal meningitis (~25-30%), because the mortality rate itself has been shown to be of significant importance for the efficacy of anti-inflammatory therapy in experimental sepsis models (Eichacker et al 2002). A beneficial effect of anti-inflammatory treatment in experimental sepsis was only observed with high mortality rates, when high inoculum sizes were used, in contrast to a harmful effect observed in clinical trials with a lower mortality rate. A likely explanation could be that high inoculum sizes induce an overwhelming inflammatory burst (e.g. high CSF cytokine levels), as shown in pneumococcal meningitis as discussed above, which may be an "artificial" situation, where anti-inflammatory treatment in particular may have a beneficial effect. 3) Adult and neonatal animal models may provide conflicting results due to difference in susceptibility to the infection (e.g. neonatal neurons become more likely apoptotic than adult neurons during oxidative stress). Thus, the choice of animal model and endpoint parameters may have significant impact on the results obtained experimentally.

In our laboratory, we have established an adult rat model using inoculum sizes of $\sim 1 \times 10^6$ CFU live pneumococci for studying outcome (Brandt et al 2004), because we believe that this model closely resembles human pneumococcal meningitis. Adjunctive therapy should be initiated during fully developed meningitis and at the time of initiation of antibiotic therapy, when studying the efficacy of adjunctive therapy (e.g. anti-inflammatory treatment). Conversely, experimental designs with initiation of adjunctive therapy at time of infection or with the use of gene-modified animals may be useful for the characterisation of important mediators in the pathophysiology of meningitis, but does not reflect treatment efficacy in a "realistic" situation that may lead to further clinical testing. In conclusion, experimental studies with pharmacological intervention or the use of gene-modified animals in pneumococcal meningitis have shown that anti-inflammatory therapy has detrimental effects in pneumococcal meningitis (Table 2): 1) A reduction in pleocytosis or intracerebral complication by anti-inflammatory therapy may be associated with increased systemic complication due to an impaired host response that may result in a poorer outcome (Brandt et al 2005; Koedel et al 2004). 2) The efficacy of adjunctive therapy may depend on disease stage or bacterial pathogens (Leib et al 1996a), i.e. an inhibition of the vasodilator NO was beneficial in some studies (Koedel et al 1995; Koedel et al 2001), but harmful in other studies (Leib et al 1998; Winkler et al 2001). 3) Hippocampal injury may have another pathogenesis than other IC complications, since therapy with dexamethasone aggravates neural apoptosis in the hippocampus (Leib et al 2003), but reduces other intracranial complications (Koedel et al 1994). Thus, testing of various anti-inflammatory interventions should optimally be addressed in various animal models before further clinical trials.

11. CONCLUSION

In the clinical studies, we found that the most common predisposing focus was otitis media in both adults and children (~30% of cases), and our results suggest that inadequate antibiotic therapy of otitis media may be associated with the development of pneumococcal meningitis. Of the classic signs of meningitis, fever and decreased consciousness were found in almost all patients with pneumococcal meningitis, whereas back rigidity was found only in half the cases, suggesting that meningitis also should be considered in unconscious febrile patients without back rigidity. We found that the case fatality rate of pneumococcal meningitis in Denmark has not improved significantly over half a century, and the death of the patient was due to both neurological and systemical complications. Risk factors for a poor clinical outcome of pneumococcal meningitis were advanced age, the predisposing focus of the infection, the infecting serotype, having a CT-scan prior to lumbar puncture, development of adjacent complications (e.g. convulsion, respiratory insufficiency) and various CSF alterations (low WBC, high protein levels, low CSF/blood glucose ratio), respectively. CSF analysis showed that IL-8 to some degree was useful as diagnostic marker in patients suspected of having meningitis on admission, and YKL-40 and sUPAR were useful as prognostic markers. However, no single CSF parameter alone distinguishes bacterial from viral meningitis or predicts the clinical outcome.

In the experimental studies, we showed that several factors influenced the CSF pleocytosis: 1) the infecting pathogen, 2) blocking of adhesion molecules (the selectin-blocker, fucoidin), 3) inhibition of chemotactic signal (IL-8), 4) inhibition of chemotactic ability of blood leukocytes (G-CSF-pretreatment) and 5) reducing the number of peripheral leukocytes (induced by early onset bacteraemia). Moreover, we documented an accelerated CSF IL-8 production with attenuation of leukocyte entry into the CSF suggesting a local release of IL-8 in the CSF during pneumococcal meningitis. Furthermore, serotype-related differences in histopathological alterations were found, indicating that the pathophysiology may vary according to the serotype. Finally, we showed that moxifloxacin had an excellent CSF penetration through inflamed and uninflamed meninges and excellent bacterial killing suggesting it to be a promising candidate for treatment of bacterial meningitis.

12. PERSPECTIVE

Because the case fatality rate has not decreased significantly over decades, further studies on pneumococcal meningitis are still warranted. The following issues should be addressed in future studies:

A better understanding of risk factors for developing meningitis and for the outcome. The analysis of genetic polymorphism from meningitis cases could identify groups of patients with a high risk for developing meningitis or with a high case fatality rate, and the analysis of pneumococcal isolates from meningitis patients could help identifying other virulence factors than the serotype of importance for both developing meningitis and mortality. Such studies could help the designing of new vaccines and the identification of patients groups with a specific need for vaccination programs. Also, experimental studies using gene-modified animals and genetically modulated pneumococcal mutants could be beneficial in addressing the interaction between various pneumococcal virulence factors and host immune reactions. Finally, international studies on pneumococcal meningitis due to different predisposing infections (e.g. otitis media) could help identifying optimal antibiotic treatment regimes for such infections, which in turn may reduce the risk of developing meningitis.

Improvements in therapy of systemic complications in pneumococcal meningitis. Because approximately half the deaths from pneumococcal meningitis are due to systemic complications, experimental studies addressing this issue are warranted. In particular, studies investigating the effect of passive immunisation with serotype-specific antibodies could result in promising adjunctive treatment options in future clinical trials, but also experimental studies with "sepsis" doses of corticosteroids or with activated protein C would be interesting study issues.

Improvements in the control of cerebral autoregulation in pneumococcal meningitis. Loss of autoregulation and the development of brain oedema are characteristic features of pneumococcal meningitis, however little is known about this issue. Future experimental studies investigating the effect of various therapeutic interventions (e.g. hyperventilation) in restoring the cerebral autoregulation and its impact on preventing brain oedema, increased intracranial pressure, and brain damage are still needed and could lead to improved treatment options. Other potential adjunctive therapies could include erythropoietin and ventricular drainage/continous monitoring of intracranial pressure and pharmacological intervention to reduce the intracranial pressure. Table 2. Selected outcome studies of immune modulation in experimental pneumococcal meningitis.

Intervention	Animals	Outcome
Activation		
TI B-2 deficiency	Mice	[^] mortality (Echchannaoui et al 2002). [^] ICP [^] BBB permeability (Koedel et al 2003)
MyD88 deficiency	Mice	fmortality \downarrow ICP \downarrow BBB permeability (Koedel et al 2002).
NfrB1 deficiency	Mice	Amortality, Vici, V BBB permeability (Rocaci et al 2004)
NFkB inhibition (pre-treatment)	Rats	\downarrow ICP \downarrow BBB permeability (Koedel et al 2000)
	nats	
Cytokines		
TNFα-deficiency	Mice	Tmortality, THNA, TLD (Gerber et al 2004), \rightarrow mortality, \rightarrow HNA (Wellmer et al 2001)
Caspase-1 deficiency	Mice	VICP, VBBB permeability (Koedel et al 2002b)
Caspase-3 inhibition (pre-treatment)	Rats	↓HNA (Gianinazzi et al 2003)
Caspase inhibition (pre-treatment)	Rabbits	
(treatment)		\rightarrow HNA (Braun et al 1999)
IL-1 receptor deficiency	Mice	mortality (Zwijnenburg et al 2003c)
IL-6 deficiency	Mice	VICP, VBBB permeability (Paul et al 2003b)
IL-6 Inhibition (pre-treatment and treatment)	Rats	VICP (Paul et al 2003b)
IL-10 deficiency	IVIICE	→mortality (Zwijnenburg et al 2003d)
IL-10 (pre-treatment and treatment)	Rats	VICP, VBBB permeability (Koedel et al 1996)
TCF02 are treatment	Nice	Vition calley (Zwijnenburg et al 2003b)
CD05 deficiency	Rats	VICP and VBBB permeability (Prister et al 1992a)
CD93-deficiency	IVIICE	\rightarrow mortanty and \rightarrow Cr, \rightarrow BBB permeability (radi et al 2004)
Adhesion molecules		
Selectin antagonist (pre-treatment)	Rats	\uparrow mortality, \rightarrow cortical injury
(treatment)		ightarrowmortality, $ ightarrow$ cortical injury (Brandt et al 2005)
CD11/18 blockage (pre-treatment)	Rabbits	\downarrow mortality and \downarrow brain oedema (Tuomanen et al 1989), \downarrow HNA (Braun et al 1999)
ICAM-deficiency	Mice	\rightarrow HNA (Gerber et al 2001)
Leukocyte activation		
MMP inhibition (pre-treatment)	Rats	cortical pecrosis (Leib et al 2000)
MMP and TNEq converting enzyme inhibition	Rats	\downarrow mortality \downarrow cortical injury \downarrow HNA and \downarrow LD (Leib et al 2001: Meli et al 2004)
(treatment and pre-treatment)	hats	
MMP-9 deficiency	Mice	→mortality (Bottcher et al 2003a)
		······································
Antioxidants		
NAC pre-treatment	Rats	\downarrow mortality, \downarrow cortical injury (Auer et al 2000), \downarrow HL (Klein et al 2003)
MnTBAP treatment	Rats	\downarrow ICP (Kastenbauer et al 2002b), \downarrow HL (Klein et al 2003)
Urate (pre-treatment)	Rats	\downarrow ICP, \downarrow BBB permeability (Kastenbauer et al 1999)
(treatment)		→ICP (Kastenbauer et al 2002b)
PBN pre-treatment	Rats	THNA and TLD (Loeffler et al 2001)
Superoxide dismutase pre-treatment and treatment	Rats	\downarrow ICP and \downarrow brain oedema (Koedel et al 1995; Pfister et al 1990a; Pfister et al 1992c).
DFO pre-treatment	Rats	↓ICP and ↓brain oedema (Pfister et al 1992c), ↓cortical injury (Auer et al 2000)
Lipid peroxidation inhibition (pre-treatment)	Rats	↓ICP, ↓brain oedema (Lorenzl et al 1995), ↓cortical injury (Auer et al 2000)
(treatment)		↓ICP, ↓brain oedema (Lorenzi et al 1995)
Catalase pre-treatment	Rats	↓ICP and ↓brain oedema (Pfister et al 1992b)
NOS inhibition (pre-treatment)	Rats	VICP, Vorain oedema
(treatment)		mortality (Haberi et al. 1994), \downarrow ICP and \downarrow Drain oedema (Koedel et al. 1995)
eNOS deficiency	Mice	ICP, IBBB permeability, I mortality (Koedel et al 2001)
RAZeber deficiency	Mice	↓BBB permeability (winkler et al 2001) [↑] BBB permeability (Schaper et al 2002)
Crollabox deficiency	Mise	BBB permeability (Schaper et al 2003)
Boly(ADB ribose) polymorose 1 deficiency	Mice	\rightarrow BBB permeability (schaper et al 2003)
Poly(ADP-ribose) polymerase-1 deficiency	whice	
Corticosteroids		
Methylprednisolone (pre-treatment, treatment)	Rats	↓ICP, ↓brain oedema (Koedel et al 1994)
Methylprednisolone (treatment)	Rabbits	\downarrow brain oedema (Tauber et al 1985)
Dexamethasone (treatment)	Rats	↑HNA and ↑LD (Leib et al 2003)
Dexamethasone (treatment)	Rabbits	↑HNA (Zysk et al 1996), \downarrow HL (Bhatt et al 1995), \downarrow brain oedema, \downarrow ICP (Tauber et al 1985)
Neuroexitation		
Glutamine synthetase inhibition (treatment)	Rabbits	↑HNA (Tumani et al 2000)
Glutamate receptor antagonist (pre-treatment)	Rats	\rightarrow HNA (Kolarova et al 2003)
Autoropylation		
Autoregulation	Data	
Endothelin-B-receptor deficiency	Rats	HNA (Ehrenreich et al 2000)
Endothelin antagonist (pre-treatment)	Rats	\downarrow cortical injury (Pfister et al 2000) and \downarrow ICP, \downarrow brain oedema (Koedel et al 1998)
Bradykinin receptor antagonist (treatment)	Rats	↓ICP and ↓brain oedema (Lorenzi et al 1996)
Other types of immune modulation		
UPAR- and tPA-deficiency	Mice	\rightarrow ICP, \rightarrow BBB permeability (Paul et al 2005).
M-CSF deficiency	Mice	\rightarrow HNA (Gerber et al 2001)
G-CSF (pre-treatment)	Rats	\downarrow mortality and \downarrow cortical injury (Brandt et al 2004), \uparrow HL (Brandt et al 2006a)
(treatment)		ightarrowmortality and $ ightarrow$ cortical injury (Brandt et al 2004), $ ightarrow$ HL (Brandt et al 2006a)
lpha-melatonin stimulating hormone (treatment)	Rats	\rightarrow ICP, \rightarrow BBB permeability (Kastenbauer et al 2001)
Melatonin pre-treatment	Rabbits	\downarrow HNA (Gerber et al 2005)
Pentoxifylline treatment	Rabbits	→HNA (Zysk et al 1997a)
Indomethacin (pre-treatment)	Rabbits	↓brain oedema (Tureen et al 1991)
Indomethacin (pre-treatment)	Rats	↓brain oedema (Pfister et al 1990a)
Bcl-2 deficiency	Mice	↑mortality, \rightarrow HNA (Wellmer et al 2004)

A better understanding of the pathophysiological events leading to hearing loss in pneumococcal meningitis. The relative significance of the meningeal pathogen and host inflammatory reactions for the development of hearing loss in pneumococcal meningitis are not fully elucidated. Serotype-related differences or other pneumococcal virulence factors may be of importance, and anti-inflammatory treatment (e.g. blocking of leukocyte recruitment) could be a beneficial treatment option.

Introduction of new antibiotics with a favourable pharmacodynamic profile (e.g. causing less inflammation) in the treatment of pneumococcal meningitis. Both experimental and clinical trials are needed to investigate the treatment effect of new antibiotics. In particular, antibiotics inhibiting bacterial protein synthesis should be tested (e.g. fluoroquinolones, clindamycin, and rifampicin).

Use of non-invasive techniques in experimental pneumococcal meningitis. Non-invasive techniques (e.g. bioluminescence) may be important tools in studying the pathogenesis and pathophysiology of pneumococcal meningitis. Also, improvements in MRI-techniques make it possible to monitor disease progress (e.g. blood/brain barrier alterations, cerebral oedema, cerebral blood flow, brain damage) over time in each individual animal. Moreover, marking pneumococci with iron, plasmids etc. – possible to visualise by non-invasive techniques – could give information of the in vivo distribution of pneumococci during the course of meningitis. Furthermore, MRI can detect fluoromolecules and thereby visualise in vivo distribution of fluoroquinolones and interaction with bacteria.

International cooperation between meningitis groups. International cooperation between meningitis groups will be beneficial for future testing of new treatments regimes in pneumococcal meningitis, because the effect of anti-inflammatory therapy may vary between experimental meningitis models.

13. SUMMARY

This thesis is based on the work performed during a 10-year period at Statens Serum Institut and Department of Infectious Diseases, Hvidovre Hospital. The investigations were performed as both clinical and experimental studies.

Based on national registration of pneumococcal meningitis cases, we showed in clinical studies that the incidence of pneumococcal meningitis was $\sim 2/100.000$ cases per year, and the most common predisposing infection focus was otitis media in ~30% of cases, results comparable with previous Danish studies. Our results suggest that inadequate antibiotic therapy of otitis media may be associated with the development of pneumococcal meningitis. Fever and decreased consciousness were found in almost all patients with pneumococcal meningitis, whereas the classical meningitis sign back rigidity was found in 1 out of every 2 cases, suggesting that meningitis also should be considered in unconscious febrile patients without back rigidity. We found that the case fatality rate of pneumococcal meningitis in Denmark has not improved significantly over half a century and remains as high as $\sim 21\%$ (adults: $\sim 30\%$ and children: ~5%) and was due to both neurological and systemical complications. Risk factors for a poor clinical outcome of pneumococcal meningitis were advanced age, the predisposing focus of the infection, the infecting serotype, having a CT-scan prior to lumbar puncture, development of adjacent complications (e.g. convulsion, respiratory insufficiency) and various CSF alterations (low WBC, high protein levels, low CSF/blood glucose ratio).

CSF samples obtained from patients admitted to Department of Infectious Diseases, Copenhagen University Hospital Hvidovre suspected for meningitis were analysed for the content of various inflammatory markers and correlated to clinical data to describe, whether such analyses could have diagnostic and prognostic value. Our results showed that the chemotactic factor IL-8 was highly elevated in patients with meningitis and that IL-8 to some degree was useful as diagnostic marker. YKL-40 and sUPAR, inflammatory prognostic markers in other infections and cancer diseases, were elevated in CSF from meningitis patients and were to some degree useful as prognostic markers. However no single CSF parameter alone distinguishes bacterial from viral meningitis or predicts the clinical outcome.

In experimental studies using a rabbit meningitis model, we studied various mechanism leading to the influx of leukocytes into the CSF and found that several factors influenced the CSF pleocytosis: 1) the infecting pneumococcal serotype, 2) blocking of adhesion molecules (the selectin-blocker fucoidin), 3) inhibition of chemotactic signal (IL-8), 4) inhibition of chemotactic ability of blood leukocytes (G-CSF-pretreatment), and 5) decrease in number of peripheral leukocytes due to early onset bacteraemia. We documented an accelerated CSF IL-8 production associated with an attenuated pleocytosis indicating local release of IL-8 into the CSF. Also, we used the rabbit meningitis model for pharmacokinetic and pharmacodynamic studies of antibiotics. We showed that moxifloxacin had an excellent CSF penetration through inflamed and uninflamed meninges and caused an excellent bacterial killing suggesting it to be a promising candidate for treatment of bacterial meningitis.

Because the rabbit model is not suitable for investigating outcome (mortality, sequelae, brain damage), we established a rat meningitis model and found that the degree and pattern of brain damage was dependent on the infecting pneumococcal serotype. This model is excellent to investigate the efficacy of various anti-inflammatory treatments and other therapeutic regimes.

Future progress may rather come from prevention (e.g. vaccination) and early identification and treatment of disposing factors. Optimal future therapeutic treatment strategies should not only involve therapy against intracranial complications but also against systemic complications.

ABBREVIATIONS

LD:	Learning deficit
CSF:	Cerebrospinal fluid
CNS:	Central nervous system
CT:	Computer tomography
MR:	Magnetic resonance
WBC:	White blood cell
IL:	Interleukin
TNF:	Tumour necrosis factor
LPS:	Lipopolysaccharide
BBB:	Blood brain barrier
OR:	Odds ratio
MIC:	Minimal inhibitory concentration
MBC:	Minimal bactericidal concentration
$T_{>MIC}$:	Duration of the concentration curve above MIC
AUC:	Area under the concentration curve
CFU:	Colony forming units
HNA:	Hippocampal neural apoptosis
HL:	Hearing loss
BBB:	Blood/brain barrier
ICP:	Intracranial pressure

References

- Ahmed, A., Jafri, H., Lutsar, I., McCoig, C. C., Trujillo, M., Wubbel, L., Shelton, S., and McCracken, G. H., Jr. (1999). Pharmacodynamics of vancomycin for the treatment of experimental penicillin- and cephalosporinresistant pneumococcal meningitis. Antimicrob Agents Chemother 43, 876-881.
- Andersson, P. B., Perry, V. H., and Gordon, S. (1992). Intracerebral injection of proinflammatory cytokines or leukocyte chemotaxins induces minimal myelomonocytic cell recruitment to the parenchyma of the central nervous system. J.Exp.Med. 176, 1, 255-259.
- Angstwurm, K., Weber, J. R., Segert, A., Burger, W., Weih, M., Freyer, D., Einhaupl, K. M., and Dirnagl, U. (1995). Fucoidin, a polysaccharide inhibiting leukocyte rolling, attenuates inflammatory responses in experimental pneumococcal meningitis in rats. Neurosci.Lett. 191, 1-4.
- Annane, D., Sebille, V., Charpentier, C., Bollaert, P. E., Francois, B., Korach, J. M., Capellier, G., Cohen, Y., Azoulay, E., Troche, G., Chaumet-Riffaut,

P., and Bellissant, E. (2002). Effect of treatment with low doses of hydrocortisone and fludrocortisone on mortality in patients with septic shock. JAMA 288, 7, 862-871.

Appelbaum, E. and Nelson, J. (1945). Penicillin in the treatment of pneumococcic meningitis. JAMA 128, 778-781.

- Aronin, S. I., Peduzzi, P., and Quagliarello, V. J. (1998). Community-acquired bacterial meningitis: Risk stratification for adverse clinical outcome and effect of antibiotic timing. Ann Intern.Med 129, 11, 862-869.
- Ashwal, S., Perkin, R. M., Thompson, J. R., Schneider, S., and Tomasi, L. G. (1994). Bacterial meningitis in children: current concepts of neurologic management. Curr.Probl.Pediatr. 24, 8, 267-284.
- Auburtin, M., Porcher, R., Bruneel, F., Scanvic, A., Trouillet, J. L., Bedos, J. P., Regnier, B., and Wolff, M. (2002). Pneumococcal meningitis in the intensive care unit: prognostic factors of clinical outcome in a series of 80 cases. Am.J Respir.Crit Care Med. 165, 5, 713-717.
- Auer, M., Pfister, L. A., Leppert, D., Tauber, M. G., and Leib, S. L. (2000). Effects of clinically used antioxidants in experimental pneumococcal meningitis. J.Infec.Dis. 182, 1, 347-350.
- Austrian, R. (1981a). Pneumococcus: the first one hundred years. Rev.Infect.Dis. 3, 2, 183-189.
- Austrian, R. (1981b). Some observations on the pneumococcus and on the current status of pneumococcal disease and its prevention. Rev.Infect.Dis. 3 Suppl, S1-17.
- Avery, O. T., Chickering, H. T., Cole, R., and Dochez, A. R (1917). Acute lobar pneumonia. Prevention and serum treatment. Monograph of the Rockefeller Institute for Medical Research, no.7
- Baird, D. R., Whittle, H. C., and Greenwood, B. M. (1976). Mortality from pneumococcal meningitis. Lancet 2, 7999, 1344-1346.
- Bhatt, S. M., Cabellos, C., Nadol, J. B J., Halpin, C., Lauretano, A., Xu, W. Z., and Tuomanen, E. (1995). The impact of dexamethasone on hearing loss in experimental pneumococcal meningitis. Pediatr.Infect.Dis.J. 14, 93-96.
- Bhatt, S. M., Lauretano, A., Cabellos, C., Halpin, C., Levine, R. A., Xu, W. Z., Nadol, J. B J., and Tuomanen, E. (1993). Progression of hearing loss in experimental pneumococcal meningitis: correlation with cerebrospinal fluid cytochemistry. J.Infect.Dis. 167, 675-683.
- Bitsch, A., Trostdorf, F., Bruck, W., Schmidt, H., Fischer, F. R., and Nau, R. (1997). Central nervous system TNF alpha-mRNA expression during rabbit experimental pneumococcal meningitis. Neurosci Lett 237, 2-3, 105-108.
- Blasi, F. and Carmeliet, P. (2002). uPAR: A versatile signalling orchestator. Nature 3, 932-943.
- Bohr, V., Rasmussen, N., Hansen, B., Gade, A., Kjersem, H., Johnsen, N., and Paulson, O. (1985). Pneumococcal meningitis: an evaluation of prognostic factors in 164 cases based on mortality and on a study of lasting sequelae. Jornal of Infection 10, 2, 143-157.
- Bonsu, B. K. and Harper, M. B. (2001). Fever interval before diagnosis, prior antibiotic treatment, and clinical outcome for young children with bacterial meningitis. Clin.Infect.Dis. 32, 4, 566-572.
- Borregaard, N. (1996). Den neutrofile granulocyts cellebiologi. Ugeskrift for Læger 158, 3908-3912.
- Bottcher, T., Gerber, J., Wellmer, A., Smirnov, A. V., Fakhrjanali, F., Mix, E., Pilz, J., Zettl, U. K., and Nau, R. (2000). Rifampin reduces production of reactive oxygen species of cerebrospinal fluid phagocytes and hippocampal neuronal apoptosis in experimental Streptococcus pneumoniae meningitis. J.Infect.Dis. 181, 6, 2095-2098.
- Bottcher, T., Ren, H., Goiny, M., Gerber, J., Lykkesfeldt, J., Kuhnt, U., Lotz, M., Bunkowski, S., Werner, C., Schau, I., Spreer, A., Christen, S., and Nau, R. (2004). Clindamycin is neuroprotective in experimental Streptococcus pneumoniae meningitis compared with ceftriaxone. J Neurochem. 91, 6, 1450-1460.
- Bottcher, T., Spreer, A., Azeh, I., Nau, R., and Gerber, J. (2003a). Matrix metalloproteinase-9 deficiency impairs host defense mechanisms against Streptococcus pneumoniae in a mouse model of bacterial meningitis. Neurosci.Lett. 338, 3, 201-204.
- Bottcher, T., von Mering, M., Ebert, S., Meyding-Lamade, U., Kuhnt, U., Gerber, J., and Nau, R. (2003b). Differential regulation of Toll-like receptor mRNAs in experimental murine central nervous system infections. Neurosci.Lett. 344, 1, 17-20.
- Brandt, C. T., Caye-Thomassen, P., Lund, S. P., Worsøe, L., Østergaard, C., Frimodt-Moller, N., Espersen, F., Thomsen, J., and Lundgren, J. D. (2006a). Hearing loss and cloclear damage in experimental pneumococcal meningitis, with special reference to the role of neutrophil granulocytes. Neurobiol.Dis. 23, 2, 300-311.
- Brandt, C. T., Lundgren, J. D., Frimodt-Moller, N., Christensen, T., Benfield, T., Espersen, F., Hougaard, D. M., and Østergaard, C. (2005). Blocking of leukocyte accumulation in the cerebrospinal fluid augments bacteremia and increases lethality in experimental pneumococcal meningitis. J Neuroimmunol. 166, 1-2, 126-131.
- Brandt, C. T., Lundgren, J. D., Lund, S. P., Frimodt-Møller, N., Christensen, T., Espersen, F., Hovgaard, D., and Østergaard, C. (2004). Attenuation of the bacterial load in blood by pretreatment with granulocyte-colonystimulating factor protects rats from fatal outcome and brain damage dur-

ing Streptococcus pneumoniae meningitis. Infection and Immunity 72, 8, 4647-4653.

- Brandt, C. T., Østergaard, C., Lundgren, J. D., and Rowland, I. J. (2006b). In vivo study of experimental pneumococcal meningitis using Magnetic Resonance Imaging. Neurobiol.Dis (submitted),
- Braun, J. S., Novak, R., Herzog, K. H., Bodner, S. M., Cleveland, J. L., and Tuomanen, E. I. (1999). Neuroprotection by a caspase inhibitor in acute bacterial meningitis. Nat.Med. 5, 3, 298-302.
- Braun, J. S., Sublett, J. E., Freyer, D., Mitchell, T. J., Cleveland, J. L., Tuomanen, E. I., and Weber, J. R. (2002). Pneumococcal pneumolysin and H(2)O(2) mediate brain cell apoptosis during meningitis. J Clin.Invest 109, 1, 19-27.
- Cairns, H. and Russell, D. S. (1946). Cerebral arteritis and phlebitis in pneumococcal meningitis. Journal Pathology and Bacteriology 58, 649-665. Casado-Flores, J., Aristegui, J., de Liria, C. R., Martinon, J. M., and Fernan-
- Casado-Flores, J., Aristegui, J., de Liria, C. R., Martinon, J. M., and Fernandez, C. (2005). Clinical data and factors associated with poor outcome in pneumococcal meningitis. Eur.J.Pediatr.1-5.
- Chavanet, P., Bonnotte, B., Guiguet, M., Zeller, V., Solary, E., Maurice, L., Casasnovas, O., Caillot, D., Waldner, A., Kisterman, J. P., and et-al (1992). High concentrations of intrathecal interleukin-6 in human bacterial and nonbacterial meningitis. J.Infect.Dis. 166, 428-431.
- Cherian, T., Lalitha, M. K., Manoharan, A., Thomas, K., Yolken, R. H., and Steinhoff, M. C. (1998). PCR-enzyme immunoassay for detection of Streptococcus pneumoniae DNA in cerebrospinal fluid samples from patients with culture-negative meningitis. J Clin.Microbiol. 36, 12, 3605-3608.
- Cintin, C., Johansen, J. S., Christensen, I. J., Price, P. A., Sçrensen, S., and Nielsen, H. J. (1999). Serum YKL-40 and colorectal cancer. Br.J.Cancer 79, 9-10, 1494-1499.
- Corless, C. E., Guiver, M., Borrow, R., EdwardsJones, V., Fox, A. J., and Kaczmarski, E. B. (2001). Simultaneous detection of Neisseria meningitidis, Haemophilus influenzae, and Streptococcus pneumoniae in suspected cases of meningitis and septicemia using real- time PCR. J.Clin.Microbiol. 39, 4, 1553-1558.
- Cottagnoud, P. H. and Tauber, M. G. (2004). New therapies for pneumococcal meningitis. Expert.Opin.Investig.Drugs 13, 4, 393-401.
- Crosson, F. J., Jr., Winkelstein, J. A., and Moxon, E. R. (1976). Participation of complement in the nonimmune host defense against experimental Haemophilus influenzae type b septicemia and meningitis. Infection and Immunity 14, 4, 882-887.
- Cundell, D. R., Gerard, N. P., Gerard, C., Idanpaan-Heikkila, I., and Tuomanen, E. I. (1995a). Streptococcus pneumoniae anchor to activated human cells by the receptor for platelet-activating factor. Nature 377, 435-438.
- Cundell, D. R., Weiser, J. N., Shen, J., Young, A., and Tuomanen, E. I. (1995b). Relationship between colonial morphology and adherence of Streptococcus pneumoniae. Infection and Immunity 63, 757-761.
- Cutts, F. T., Zaman, S. M., Enwere, G., Jaffar, S., Levine, O. S., Okoko, J. B., Oluwalana, C., Vaughan, A., Obaro, S. K., Leach, A., McAdam, K. P., Biney, E., Saaka, M., Onwuchekwa, U., Yallop, F., Pierce, N. F., Greenwood, B. M., and Adegbola, R. A. (2005). Efficacy of nine-valent pneumococcal conjugate vaccine against pneumonia and invasive pneumococcal disease in The Gambia: randomised, double-blind, placebo-controlled trial. Lancet 365, 9465, 1139-1146.
- Dacey, R. G. and Sande, M. A. (1974). Effect of probenecid on cerebrospinal fluid concentrations of penicillin and cephalosporin derivatives. Antimicrobial Agents and Chemotherapy 6, 437-441.Dagan, R., Givon-Lavi, N., Zamir, O., Sikuler-Cohen, M., Guy, L., Janco, J.,
- Dagan, R., Givon-Lavi, N., Zamir, O., Sikuler-Cohen, M., Guy, L., Janco, J., Yagupsky, P., and Fraser, D. (2002). Reduction of nasopharyngeal carriage of Streptococcus pneumoniae after administration of a 9-valent pneumococcal conjugate vaccine to toddlers attending day care centers. J Infect Dis 185, 7, 927-936.
- de Gans, J. and van de Beek, D. (14-11-2002). Dexamethasone in adults with bacterial meningitis. N.Engl.J.Med. 347, 20, 1549-1556.
- DelMaschio, A., DeLuigi, A., MartinPadura, I., Brockhaus, M., Bartfai, T., Fruscella, P., Adorini, L., Martino, G. V., Furlan, R., DeSimoni, M. G., and Dejana, E. (1999). Leukocyte recruitment in the cerebrospinal fluid of mice with experimental meningitis is inhibited by an antibody to junctional adhesion molecule (JAM). J.Exp.Med. 190, 9, 1351-1356.
- Deng, G. M., Liu, Z. Q., and Tarkowski, A. (2001). Intracisternally localized bacterial DNA containing CpG motifs induces meningitis. J Immunol. 167, 8, 4616-4626.
- Diab, A., Zhu, J., Lindquist, L., Wretlind, B., Bakhiet, M., and Link, H. (1997). Haemophilus influenzae and Streptococcus pneumoniae induce different intracerebral mRNA cytokine patterns during the course of experimental bacterial meningitis. Clin Exp Immunol 109, 2, 233-241.
- Djukic, M., Bottcher, T., Wellmer, A., Gerber, J., Brocke, V. V., Eiffert, H., and Nau, R. (2005). Moxifloxacin in experimental Streptococcus pneumoniae cerebritis and meningitis. Neurocrit.Care 2, 3, 325-329.
- Dumont, R. A., Car, B. D., Voitenok, N. N., Junker, U., Moser, B., Zak, O., and OReilly, T. (2000). Systemic neutralization of interleukin-8 markedly reduces neutrophilic pleocytosis during experimental lipopolysaccharideinduced meningitis in rabbits. Infection and Immunity 68, 5756-5763.

- Echchannaoui, H., Frei, K., Schnell, C., Leib, S. L., Zimmerli, W., and Landmann, R. (2002). Toll-like receptor 2-deficient mice are highly susceptible to Streptococcus pneumoniae meningitis because of reduced bacterial clearing and enhanced inflammation. J Infect. Dis. 186, 6, 798-806.
- Ehrenreich, H., Nau, R., Dembowski, C., Hasselblatt, M., Barth, M., Hahn, A., Schilling, L., Siren, A. L., and Bruck, W. (2000). Endothelin B receptor deficiency is associated with an increased rate of neuronal apoptosis in the dentate gyrus. Neuroscience 95, 4, 993-1001.
- Eichacker, P. Q., Parent, C., Kalil, A., Esposito, C., Cui, X., Banks, S. M., Gerstenberger, P., Fitz, Y., Danner, R. L., and Natanson, C. (2002). Risk and the Efficacy of Antiinflammatory Agents: Retrospective and Confirmatory Studies of Sepsis. American Journal of Respiratory and Critical Care Medicine 166, 9, 1197-1205.
- Eriksson, M., Henriques, B., and Ekdahl, K. (2000). Epidemiology of pneumococcal infections in Swedish children. Acta Paediat. 89:35-39, 35-39.
- Ernst, J. D., Decazes, J. M., and Sande, M. A. (1983). Experimental pneumococcal meningitis: role of leukocytes in pathogenesis. Infection and Immunity 41, 275-279.
- Eskola, J., Kilpi, T., Palmu, A., Jokinen, J., Haapakoski, J., Herva, E., Takala, A., Kayhty, H., Karma, P., Kohberger, R., Siber, G., Makela, P. H., Lockhart, S., and Ecrola, M. (2001). Efficacy of a pneumococcal conjugate vaccine against acute otitis media. N.Engl.J.Med. 344, 6, 403-409.
- Fassbender, K., Mielke, O., Bertsch, T., Muehlhauser, F., Hennerici, M., Kurimoto, M., and Rossol, S. (1999). Interferon-gamma-inducing factor (IL-18) and interferon-gamma in inflammatory CNS diseases. Neurology 53, 5, 1104-1106.
- Fassbender, K., Schminke, U., Ries, S., Ragoschke, A., Kischka, U., Fatar, M., and Hennerici, M. (1997). Endothelial-derived adhesion molecules in bacterial meningitis: association to cytokine release and intrathecal leukocyte-recruitment. J.Neuroimmunol. 74, 1-2, 130-134.
- Feldman, W. E. (1977). Relation of the Concentrations of Bacteria and Bacterial Antigen in Cerebrospinal Fluid to Prognosis in Patients with Bacterial Meningitis. Medical Intelligence 296, 433-435.
- Finland, M., Brown, J. W., and Rauh, A. E. (1938). Treatment of Pneumococcic Meningitis. N.Engl.J.Med. 218, 25, 1033-1044.
- Fiore, A. E., Moroney, J. F., Farley, M. M., Harrison, L. H., Patterson, J. E., Jorgensen, J. H., Cetron, M., Kolczak, M. S., Breiman, R. F., and Schuchat, A. (2000). Clinical outcomes of meningitis caused by Streptococcus pneumoniae in the era of antibiotic resistance. Clin.Infect.Dis. 30, 1, 71-77.
- Förderreuther, S., Tatsch, K., Einhäupl, K. M., and Pfister, H. W. (1992). Abnormalities of cerebral blood flow in the acute phase of bacterial meningitis in adults. J.Neurol. 239, 8, 431-436.
- Fraser, D. W., Darby, C. P., Koehler, R. E., Jacobs, C. F., and Feldman, R. A. (1973). Risk factors in bacterial meningitis: Charleston County, South Carolina. J.Infect.Dis. 127, 3, 271-277.
- Frei, K., Nadal, D., Pfister, H. W., and Fontana, A. (1993). Listeria meningitis: identification of a cerebrospinal fluid inhibitor of macrophage listericidal function as interleukin 10. J.Exp.Med. 178, 1255-1261.
- Friedland, I. R., Paris, M. M., Hickey, S., Shelton, S., Olsen, K., Paton, J. C., and McCracken, G. H., Jr. (1995). The limited role of pneumolysin in the pathogenesis of pneumococcal meningitis. J.Infect.Dis. 172, 805-809.
- Geiseler, P. J., Nelson, K. E., Levin, S., Reddi, K. T., and Moses, V. K. (1980). Community-acquired purulent meningitis: a review of 1,316 cases during the antibiotic era, 1954-1976. Rev.Infect.Dis. 2, 5, 725-745.
- Gerber, J., Bottcher, T., Hahn, M., Siemer, A., Bunkowski, S., and Nau, R. (2004). Increased mortality and spatial memory deficits in TNF-alpha-deficient mice in ceftriaxone-treated experimental pneumococcal meningitis. Neurobiology of Disease 16, 1, 133-138.
- Gerber, J., Lotz, M., Ebert, S., Kiel, S., Huether, G., Kuhnt, U., and Nau, R. (2005). Melatonin is neuroprotective in experimental Streptococcus pneumoniae meningitis. J Infect Dis 191, 5, 783-790.
- Gerber, J., Pohl, K., Sander, V., Bunkowski, S., and Nau, R. (2003). Rifampin followed by ceftriaxone for experimental meningitis decreases lipoteichoic Acid concentrations in cerebrospinal fluid and reduces neuronal damage in comparison to ceftriaxone alone. Antimicrobial Agents and Chemotherapy 47, 4, 1313-1317.
- Gerber, J., Raivich, G., Wellmer, A., Noeske, C., Kunst, T., Werner, A., Bruck, W., and Nau, R. (2001). A mouse model of Streptococcus pneumoniae meningitis mimicking several features of human disease. Acta Neuropathol. 101, 5, 499-508.
- Gianinazzi, C., Grandgirard, D., Imboden, H., Egger, L., Meli, D. N., Bifrare, Y. D., Joss, P. C., Tauber, M. G., Borner, C., and Leib, S. L. (2003). Caspase-3 mediates hippocampal apoptosis in pneumococcal meningitis. Acta Neuropathol. (Berl) 105, 5, 499-507.
- Gillespie, S. H. and Balakrishnan, I. (2000). Pathogenesis of pneumococcal infection. J.Med.Microbiol. 49, 12, 1057-1067.
- Goetghebuer, T., West, T. E., Wermenbol, V., Cadbury, A. L., Milligan, P., Lloyd-Evans, N., Adegbola, R. A., Mulholland, E. K., Greenwood, B. M., and Weber, M. W. (2000). Outcome of meningitis caused by Streptococcus pneumoniae and Haemophilus influenzae type b in children in The Gambia. Trop.Med.Int.Health 5, 3, 207-213.
- Gordon, S. B., Walsh, A. L., Chaponda, M., Gordon, M. A., Soko, D.,

Mbwvinji, M., Molyneux, M. E., and Read, R. C. (2000). Bacterial meningitis in Malawian adults: pneumococcal disease is common, severe, and seasonal. Clin.Infect.Dis. 31, 1, 53-57.

- Grande, P. O., Myhre, E. B., Nordstrom, C. H., and Schliamser, S. (2002). Treatment of intracranial hypertension and aspects on lumbar dural puncture in severe bacterial meningitis. Acta Anaesthesiol.Scand. 46, 3, 264-270.
- Granert, C., Raud, J., and Lindquist, L. (1998). The polysaccharide fucoidin inhibits the antibiotic- induced inflammatory cascade in experimental pneumococcal meningitis. Clin.Diagn.Lab.Immunol 5, 3, 322-324.
- Granert, C., Raud, J., Waage, A., and Lindquist, L. (1999). Effects of polysaccharide fucoidin on cerebrospinal fluid interleukin- 1 and tumor necrosis factor alpha in pneumococcal meningitis in the rabbit. Infection and Immunity 67, 2071-2074.
- Granert, C., Raud, J., Xie, X., Lindquist, L., and Lindbom, L. (1994). Inhibition of leukocyte rolling with polysaccharide fucoidin prevents pleocytosis in experimental meningitis in the rabbit. J.Clin.Invest. 93, 3, 929-936.
- Graur, D., Duret, L., and Gouy, M. (1996). Phylogenetic position of the order Lagomorpha (rabbits, hares and allies). Nature 379, 6563, 333-335.
- Gray, L. D. and Fedorko, D. P. (1992). Laboratory diagnosis of bacterial meningitis. Clin.Microbiol Rev. 5, 130-145.
- Guerra-Romero, L., Tureen, J. H., Fournier, M. A., Makrides, V., and Tauber, M. G. (1993). Amino acids in cerebrospinal and brain interstitial fluid in experimental pneumococcal meningitis. Pediatr.Res. 33, 510-513.
- Haberl, R. L., Anneser, F., Koedel, U., and Pfister, H. W. (1994). Is nitric oxide involved as a mediator of cerebrovascular changes in the early phase of experimental pneumococcal meningitis? Neurol.Res. 16, 108-112.
- Harada, A., Sekido, N., Akahoshi, T., Wada, T., Mukaida, N., and Matsushima, K. (1994). Essential involvement of interleukin-8 (IL-8) in acute inflammation. J.Leukoc.Biol. 56, 5, 559-564.
- Harada, A., Sekido, N., Kuno, K., Akiyama, M., Kasahara, T., Nakanishi, I., Mukaida, N., and Matsushima, K. (1993). Expression of recombinant rabbit IL-8 in Escherichia coli and establishment of the essential involvement of IL-8 in recruiting neutrophils into lipopolysaccharide-induced inflammatory site of rabbit skin. Int.Immunol. 5, 6, 681-690.
- Hausdorff, W. P., Bryant, J., Kloek, C., Paradiso, P. R., and Siber, G. R. (2000). The contribution of specific pneumococcal serogroups to different disease manifestations: Implications for conjugate vaccine formulation and use, part II. Clin.Infect.Dis. 30, 1, 122-140.
- Hemmi, H., Takeuchi, O., Kawai, T., Kaisho, T., Sato, S., Sanjo, H., Matsumoto, M., Hoshino, K., Wagner, H., Takeda, K., and Akira, S. (2000). A Toll-like receptor recognizes bacterial DNA. Nature 408, 6813, 740-745.
- Henneberger, P. K., Galaid, E. I., and Marr, J. S. (1983). The descriptive epidemiology of pneumococcal meningitis in New York City. Am.J Epidemiol. 117, 4, 484-491.
- Ichiyama, T., Hayashi, T., and Furukawa, S. (1996). Cerebrospinal fluid concentrations of soluble tumor necrosis factor receptor in bacterial and aseptic meningitis. Neurology 46, 837-838.
- Ichiyama, T., Hayashi, T., Nishikawa, M., and Furukawa, S. (1997). Levels of transforming growth factor beta 1, tumor necrosis factor alpha, and interleukin 6 in cerebrospinal fluid: Association with clinical outcome for children with bacterial meningitis. Clin Infect Dis 25, 2, 328-329.
- Ichiyama, T., Nishikawa, M., Hayashi, T., Hayashi, S., Ryozawa, M., and Furukawa, S. (2000). Cerebrospinal fluid concentrations of alpha-melanocyte-stimulating hormone in bacterial and aseptic meningitis. Acta Paediat. 89, 7, 803-805.
- Jackson, L. A., Neuzil, K. M., Yu, O. C., Benson, P., Barlow, W. E., Adams, A. L., Hanson, C. A., Mahoney, L. D., Shay, D. K., and Thompson, W. W. (2003). Effectiveness of pneumococcal polysaccharide vaccine in older adults. N Engl J Med 348, 18, 1747-1755.
- Kadurugamuwa, J. L., Modi, K., Yu, J., Francis, K. P., Orihuela, C., Tuomanen, E., Purchio, A. F., and Contag, P. R. (2005). Noninvasive monitoring of pneumococcal meningitis and evaluation of treatment efficacy in an experimental mouse model. Mol.Imaging 4, 2, 137-142.
- Kaltoft, M. S., Zeuthen, N., and Konradsen, H. B. (2000). Epidemiology of invasive pneumococcal infections in children aged 0-6 years in Denmark: a 19-year nationwide surveillance study. Acta Paediatr.Suppl 89, 435, 3-10.
- Kanegaye, J. T., Soliemanzadeh, P., and Bradley, J. S. (2001). Lumbar puncture in pediatric bacterial meningitis: defining the time interval for recovery of cerebrospinal fluid pathogens after parenteral antibiotic pretreatment. Pediatrics. 108, 5, 1169-1174.
- Kastenbauer, S., Angele, B., Sporer, B., Pfister, H. W., and Koedel, U. (2005). Patterns of protein expression in infectious meningitis: a cerebrospinal fluid protein array analysis. J.Neuroimmunol. 164, 1-2, 134-139.
- Kastenbauer, S., Koedel, U., Becker, B. F., and Pfister, H. W. (2002a). Oxidative stress in bacterial meningitis in humans. Neurology 58, 2, 186-191. Kastenbauer, S., Koedel, U., Becker, B. F., and Pfister, H. W. (2002b). Pneu-
- Kastenbauer, S., Koedel, U., Becker, B. F., and Pfister, H. W. (2002b). Pneumococcal meningitis in the rat: evaluation of peroxynitrite scavengers for adjunctive therapy. Eur.J Pharmacol. 449, 1-2, 177-181.
- Kastenbauer, S., Koedel, U., Brzoska, T., Luger, T. A., and Pfister, H. W. (2001). Failure of alpha-melanocyte stimulating hormone to attenuate cerebral complications in experimental pneumococcal meningitis. J.Neuroimmunol. 116, 1, 56-61.

- Kastenbauer, S., Koedel, U., and Pfister, H. W. (1999). Role of peroxynitrite as a mediator of pathophysiological alterations in experimental pneumococcal meningitis. J.Infect.Dis. 180, 4, 1164-1170.
- Kastenbauer, S., Koedel, U., Weih, F., Ziegler-Heitbrock, L., and Pfister, H. W. (2004). Protective role of NF-kappaB1 (p50) in experimental pneumococcal meningitis. Eur.J Pharmacol. 498, 1-3, 315-318.
- Kastenbauer, S., Koedel, U., Wick, M., Kieseier, B. C., Hartung, H. P., and Pfister, H. W. (2003). CSF and serum levels of soluble fractalkine (CX3CL1) in inflammatory diseases of the nervous system. J Neuroimmunol. 137, 1-2, 210-217.
- Kastenbauer, S. and Pfister, H. W. (2003). Pneumococcal meningitis in adults: spectrum of complications and prognostic factors in a series of 87 cases. Brain 126, Pt 5, 1015-1025.
- Kellner, J. D., Scheifele, D. W., Halperin, S. A., Lebel, M. H., Moore, D., Le Saux, N., Ford-Jones, E. L., Law, B., and Vaudry, W. (2002). Outcome of penicillin-nonsusceptible Streptococcus pneumoniae meningitis: a nested case-control study. Pediatr.Infect Dis J 21, 10, 903-910.
- Kim, J. O. and Weiser, J. N. (1998). Association of intrastrain phase variation in quantity of capsular polysaccharide and teichoic acid with the virulence of Streptococcus pneumoniae. J Infect Dis 177, 2, 368-377.
- Klein, M., Koedel, U., Pfister, H. W., and Kastenbauer, S. (2003). Meningitisassociated hearing loss: Protection by adjunctive antioxidant therapy. Ann.Neurol. 54, 4, 451-458.
- Klein, M., Paul, R., Angele, B., Popp, B., Pfister, H. W., and Koedel, U. (2006). Protein expression pattern in experimental pneumococcal meningitis. Microbes.Infect.
- Koedel, U., Angele, B., Rupprecht, T., Wagner, H., Roggenkamp, A., Pfister, H. W., and Kirschning, C. J. (2003). Toll-like receptor 2 participates in mediation of immune response in experimental pneumococcal meningitis. J Immunol. 170, 1, 438-444.
- Koedel, U., Bayerlein, I., Paul, R., Sporer, B., and Pfister, H. W. (2000). Pharmacologic interference with NF-kappa B activation attenuates central nervous system complications in experimental pneumococcal meningitis. J Infect Dis 182, 1437-1445.
- Koedel, U., Bernatowicz, A., Frei, K., Fontana, A., and Pfister, H. W. (1996). Systemically (but not intrathecally) administered IL-10 attenuates pathophysiologic alterations in experimental pneumococcal meningitis. J.Immunol. 157, 11, 5185-5191.
- Koedel, U., Bernatowicz, A., Paul, R., Frei, K., Fontana, A., and Pfister, H. W. (1995). Experimental pneumococcal meningitis: cerebrovascular alterations, brain edema, and meningeal inflammation are linked to the production of nitric oxide. Ann.Neurol. 37, 313-323.
- Koedel, U., Lorenzl, S., Gorriz, C., Arendt, R. M., and Pfister, H. W. (1998). Endothelin B receptor-mediated increase of cerebral blood flow in experimental pneumococcal meningitis. J.Cereb.Blood Flow Metab. 18, 1, 67-74.
- Koedel, U., Paul, R., Winkler, F., Kastenbauer, S., Huang, P. L., and Pfister, H. W. (2001). Lack of endothelial nitric oxide synthase aggravates murine pneumococcal meningitis. J Neuropathol.Exp.Neurol. 60, 11, 1041-1050.
- Koedel, U. and Pfister, H. W. (1997). Protective effect of the antioxidant Nacetyl-L-cysteine in pneumococcal meningitis in the rat. Neurosci.Lett. 225, 1, 33-36.
- Koedel, U. and Pfister, H. W. (1999). Oxidative stress in bacterial meningitis. Brain Pathol. 9, 1, 57-67.
- Koedel, U., Pfister, H. W., and Tomasz, A. (1994). Methylprednisolone attenuates inflammation, increase of brain water content and intracranial pressure, but does not influence cerebral blood flow changes in experimental pneumococcal meningitis. Brain Res. 644, 25-31.
- Koedel, U., Rupprecht, T., Angele, B., Heesemann, J., Wagner, H., Pfister, H. W., and Kirschning, C. J. (2004). MyD88 is required for mounting a robust host immune response to Streptococcus pneumoniae in the CNS. Brain 127, Pt 6, 1437-1445.
- Koedel, U., Scheld, W. M., and Pfister, H. W. (2002a). Pathogenesis and pathophysiology of pneumococcal meningitis. Lancet Infect.Dis. 2, 12, 721-736.
- Koedel, U., Winkler, F., Angele, B., Fontana, A., Flavell, R. A., and Pfister, H. W. (2002b). Role of Caspase-1 in experimental pneumococcal meningitis: Evidence from pharmacologic Caspase inhibition and Caspase-1-deficient mice. Ann.Neurol. 51, 3, 319-329.
- Koedel, U., Winkler, F., Angele, B., Fontana, A., and Pfister, H. W. (2002c). Meningitis-associated central nervous system complications are mediated by the activation of poly(ADP-ribose) polymerase. J Cereb.Blood Flow Metab 22, 1, 39-49.
- Kolarova, A., Ringer, R., Tauber, M. G., and Leib, S. L. (2003). Blockade of NMDA receptor subtype NR2B prevents seizures but not apoptosis of dentate gyrus neurons in bacterial meningitis in infant rats. BMC.Neurosci. 4, 1, 21-
- Kolmer, J. A. (1929). Pneumococcus and streptococcus meningitis: chemotherapy and Serum therapy, with special references to newer methods. JAMA 92, 874-
- Konradsen, H. B. and Kaltoft, M. S. (2002). Invasive pneumococcal infections in Denmark from 1995 to 1999: epidemiology, serotypes, and resistance. Clin.Diagn.Lab Immunol. 9, 2, 358-365.

- Kornelisse, R. F., Hack, C. E., Savelkoul, H. F. J., Raan, T. C. T. M. V., Hop, W. C. J., vanMierlo, G., Suur, M. H., Neijens, H. J., and deGroot, R. (1997). Intrathecal production of interleukin-12 and gamma interferon in patients with bacterial meningitis. Infect Immun 65, 3, 877-881.
- Kornelisse, R. F., Hoekman, K., Visser, J. J., Hop, W. C. J., Huijmans, J. G. M., vanderStraaten, P. J. C., vanderHeijden, A. J., Sukhai, R. N., Neijens, H. J., and deGroot, R. (1996). The role of nitric oxide in bacterial meningitis in children. J Infect Dis 174, 1, 120-126.
- Kornelisse, R. F., Westerbeek, C. M., Spoor, A. B., van der, Heijde B., Spanjaard, L., Neijens, H. J., and de Groot, R. (1995). Pneumococcal meningitis in children: prognostic indicators and outcome. Clin.Infect.Dis. 21, 6, 1390-1397.
- Koskiniemi, M., Rantalaiho, T., Piiparinen, H., von Bonsdorff, C. H., Farkkila, M., Jarvinen, A., Kinnunen, E., Koskiniemi, S., Mannonen, L., Muttilainen, M., Linnavuori, K., Porras, J., Puolakkainen, M., Raiha, K., Salonen, E. M., Ukkonen, P., Vaheri, A., and Valtonen, V. (2001). Infections of the central nervous system of suspected viral origin: a collaborative study from Finland. J Neurovirol. 7, 5, 400-408.
- Kostyukova, N. N., Volkova, M. O., Ivanova, V. V., and Kvetnaya, A. S. (1995). A study of pathogenic factors of Streptococcus pneumoniae strains causing meningitis. FEMS Immunol.Med.Microbiol. 10, 2, 133-137.
- Kronborg, G., Østergaard, C., Weis, N., Nielsen, H., Obel, N., Pedersen, S. S., Price, P. A., and Johansen, J. S. (2002). The serum level of YKL-40 is elevated in patients with Streptococcus pneumoniae bacteremia and is associated with the outcome of the disease. Scand.J Infect.Dis. 34, 5, 323-326.
- Kyaw, M. H., Lynfield, R., Schaffner, W., Craig, A. S., Hadler, J., Reingold, A., Thomas, A. R., Harrison, L. H., Bennett, N. M., Farley, M. M., Facklam, R. R., Jorgensen, J. H., Besser, J., Zell, E. R., Schuchat, A., and Whitney, C. G. (2006). Effect of introduction of the pneumococcal conjugate vaccine on drug-resistant Streptococcus pneumoniae. N Engl J Med 354, 14, 1455-1463.
- La Scolea, L. J. and Dryja, D. (1984). Quantitation of bacteria in cerebrospinal fluid and blood of children with meningitis and its diagnostic significance. J Clin.Microbiol. 19, 2, 187-190.
- Larsen, C. S. and Bjerager, M. (1990). Determination of interleukin-2 (IL-2) and soluble IL-2 receptors (S-IL-2R) in serum and cerebrospinal fluid does not discriminate purulent and aseptic meningitis. Scand.J.Infect.Dis. 22, 3, 327-331.
- Laxer, R. M. and Marks, M. I. (1977). Pneumococcal meningitis in children. Am.J Dis.Child 131, 8, 850-853.
- Leib, S. L., Clements, J. M., Lindberg, R. L., Heimgartner, C., Loeffler, J. M., Pfister, L. A., Tauber, M. G., and Leppert, D. (2001). Inhibition of matrix metalloproteinases and tumour necrosis factor alpha converting enzyme as adjuvant therapy in pneumococcal meningitis. Brain 124, Pt 9, 1734-1742.
- Leib, S. L., Heimgartner, C., Bifrare, Y. D., Loeffler, J. M., and Tauber, M. G. (2003). Dexamethasone Aggravates Hippocampal Apoptosis and Learning Deficiency in Pneumococcal Meningitis in Infant Rats. Pediatr.Res. 54, 3, 353-357.
- Leib, S. L., Kim, Y. S., Black, S. M., Ferriero, D. M., and Tauber, M. G. (1996a). Detrimental effect of nitric oxide inhibition in experimental bacterial meningitis [letter]. Ann.Neurol. 39, 555-556.
- Leib, S. L., Kim, Y. S., Black, S. M., Tureen, J. H., and Tauber, M. G. (1998). Inducible nitric oxide synthase and the effect of aminoguanidine in experimental neonatal meningitis. J Infect Dis 177, 3, 692-700.
- Leib, S. L., Kim, Y. S. M., Ferriero, D. M., and Tauber, M. G. (1996b). Neuroprotective effect of excitatory amino acid antagonist kynurenic acid in experimental bacterial meningitis. J Infect Dis 173, 166-171.
- Leib, S. L., Leppert, D., Clements, J., and Tauber, M. G. (2000). Matrix metalloproteinases contribute to brain damage in experimental pneumococcal meningitis. Infection and Immunity 68, 2, 615-620.
- Leino, T., Auranen, K., Jokinen, J., Leinonen, M., Tervonen, P., and Takala, A. K. (2001). Pneumococcal carriage in children during their first two years: important role of family exposure. Pediatr.Infect Dis J 20, 11, 1022-1027.
- Lepper, M. H. and Dowling, H. F. (1951). Treatment of pneumococcic meningitis with penicillin compared with penicillin plus aureomycin. Arch.Intern.Med. 88, 489-494.
- Leppert, D., Leib, S. L., Grygar, C., Miller, K. M., Schaad, U. B., and Hollander, G. A. (2000). Matrix metalloproteinase (MMP)-8 and MMP-9 in cerebrospinal fluid during bacterial meningitis: Association with bloodbrain barrier damage and neurological sequelae. Clin Infect Dis 31, 80+-
- Lessing, M. P. A. and Bowler, I. C. J. (1996). The Value of Cerebrospinal Fluid Enrichment Culture in the Diagnosis of Acute Bacterial Meningitis. Eur J Clin Microbiol Infect Dis 15, 79-82.
- Lewczuk, P., Reiber, H., and Tumani, H. (1998). Intercellular adhesion molecule-1 in cerebrospinal fluid--the evaluation of blood-derived and brainderived fractions in neurological diseases. J Neuroimmunol. 87, 1-2, 156-161.
- Lexau, C. A., Lynfield, R., Danila, R., Pilishvili, T., Facklam, R., Farley, M. M., Harrison, L. H., Schaffner, W., Reingold, A., Bennett, N. M., Hadler, J., Cieslak, P. R., and Whitney, C. G. (2005). Changing epidemiology of inva-

sive pneumococcal disease among older adults in the era of pediatric pneumococcal conjugate vaccine. JAMA 294, 16, 2043-2051.

- Lindvall, P., Ahlm, C., Ericsson, M., Gothefors, L., Naredi, S., and Koskinen, L. O. (2004). Reducing intracranial pressure may increase survival among patients with bacterial meningitis. Clin.Infect Dis. 38, 3, 384-390.
- Loeffler, J. M., Ringer, R., Hablutzel, M., Tauber, M. G., and Leib, S. L. (2001). The free radical scavenger alpha-phenyl-tert-butyl nitrone aggravates hippocampal apoptosis and learning deficits in experimental pneumococcal meningitis. J.Infect.Dis. 183, 2, 247-252.
- Lopez-Cortes, L. F., Cruz Ruiz, M., Gomez Mateos, J., Jimenez Hernandez, D., Palomino, J., and Jimenez, E. (1993). Measurement of levels of tumor necrosis factor-alpha and interleukin-1 beta in the CSF of patients with meningitis of different etiologies: utility in the differential diagnosis. Clin.Infect.Dis. 16, 4, 534-539.
- Lorenzl, S., Koedel, U., Frei, K., Bernatowicz, A., Fontana, A., and Pfister, H. W. (1995). Protective effect of a 21-aminosteroid during experimental pneumococcal meningitis. J Infect.Dis. 172, 113-118.
- Lorenzl, S., Koedel, U., Frei, K., and Pfister, H. W. (1996). Effect of the bradykinin B-2 receptor antagonist Hoe140 in experimental pneumococcal meningitis in the rat. Eur J Pharmacol 308, 3, 335-341.
- Lukes, A., Mun-Bryce, S., Lukes, M., and Rosenberg, G. A. (1999). Extracellular matrix degradation by metalloproteinases and central nervous system diseases. Mol.Neurobiol. 19, 3, 267-284.
- Lund, E. (1970). Types of pneumococci found in blood, spinal fluid and pleural exudate during a period of 15 years (1954-1969). Acta Pathol.Microbiol Immunol.Scand.B. 78, 333-336.
- Lutsar, I., Ahmed, A., Friedland, I. R., Trujillo, M., Wubbel, L., Olsen, K., and McCracken, G. H., Jr. (1997). Pharmacodynamics and bactericidal activity of ceftriaxone therapy in experimental cephalosporin-resistant pneumococcal meningitis. Antimicrobial Agents and Chemotherapy 41, 11, 2414-2417.
- Lutsar, I., Friedland, I. R., Wubbel, L., McCoig, C. C., Jafri, H. S., Ng, W., Ghaffar, F., and McCracken, G. H., Jr. (1998). Pharmacodynamics of gatifloxacin in cerebrospinal fluid in experimental cephalosporin-resistant pneumococcal meningitis. Antimicrob.Agents Chemother. 42, 10, 2650-2655.
- McCoig, C. C., Wubbel, L., Jafri, H. S., Lutsar, I., Bastero, R., Olsen, K., Shelton, S., Friedland, I. R., and McCracken, G. H., Jr. (1999). Pharmacodynamics of trovafloxacin in experimental pneumococcal meningitis: basis for dosage selection in children with meningitis. J Antimicrob Chemother 43, 683-688.
- McIntosh, E. D., Conway, P., Willingham, J., Hollingsworth, R., and Lloyd, A. (2005). Pneumococcal pneumonia in the UK--how herd immunity affects the cost-effectiveness of 7-valent pneumococcal conjugate vaccine (PCV). Vaccine 23, 14, 1739-1745.
- McMillan, D. A., Lin, C. Y., Aronin, S. I., and Quagliarello, V. J. (2001). Community-acquired bacterial meningitis in adults: categorization of causes and timing of death. Clin.Infect Dis 33, 7, 969-975.
- Meli, D. N., Čhristen, S., Leib, S. L., and Tauber, M. G. (2002). Current concepts in the pathogenesis of meningitis caused by Streptococcus pneumoniae. Curr.Opin.Infect.Dis. 15, 3, 253-257.
- Meli, D. N., Coimbra, R. S., Erhart, D. G., Loquet, G., Bellac, C. L., Tauber, M. G., Neumann, U., and Leib, S. L. (2006). Doxycycline reduces mortality and injury to the brain and cochlea in experimental pneumococcal meningitis. Infection and Immunity 74, 7, 3890-3896.
- Meli, D. N., Loeffler, J. M., Baumann, P., Neumann, U., Buhl, T., Leppert, D., and Leib, S. L. (2004). In pneumococcal meningitis a novel water-soluble inhibitor of matrix metalloproteinases and TNF-alpha converting enzyme attenuates seizures and injury of the cerebral cortex. J Neuroimmunol. 151, 1-2, 6-11.
- Mertsola, J., Kennedy, W. A., Waagner, D., Saez-Llorens, X., Olsen, K., Hansen, E. J., and McCracken, G. H., Jr. (1991). Endotoxin concentrations in cerebrospinal fluid correlate with clinical severity and neurologic outcome of Haemophilus influenzae type B meningitis. Am.J.Dis.Child 145, 1099-1103.
- Meyer, C. N., Samuelsson, I. S., Galle, M., and Bangsborg, J. M. (2004). Adult bacterial meningitis: aetiology, penicillin susceptibility, risk factors, prognostic factors and guidelines for empirical antibiotic treatment. Clin.Microbiol.Infect 10, 8, 709-717.
- Michel, U., Ebert, S., Schneider, O., Shintani, Y., Bunkowski, S., Smirnov, A., Stringaris, A., Gerber, J., Bruck, W., and Nau, R. (2000). Follistatin (FS) in human cerebrospinal fluid and regulation of FS expression in a mouse model of meningitis. Eur.J.Endocrinol. 143, 6, 809-816.
- Mitchell, L., Smith, S. H., Braun, J. S., Herzog, K. H., Weber, J. R., and Tuomanen, E. I. (2004). Dual phases of apoptosis in pneumococcal meningitis. J Infect Dis 190, 11, 2039-2046.
- Møller, K., Høgh, P., Larsen, F. S., Strauss, G. I., Skinhøj, P., Sperling, B. K., and Knudsen, G. M. (2000a). Regional cerebral blood flow during hyperventilation in patients with acute bacterial meningitis. Clin.Physiol. 20, 5, 399-410.
- Møller, K., Larsen, F. S., Bie, P., and Skinhøj, P. (2001a). The syndrome of inappropriate secretion of antidiuretic hormone and fluid restriction in meningitis – how strong is the evidence? Scand.J Infect Dis 33, 1, 13-26.

- Møller, K., Larsen, F. S., Qvist, J., Wandall, J. H., Knudsen, G. M., Gjorup, I. E., and Skinhøj, P. (2001b). Dependency of cerebral blood flow on mean arterial pressure in patients with acute bacterial meningitis. Crit.Care Med. 28, 4, 1027-1032.
- Møller, K., Skinhøj, P., Knudsen, G. M., and Larsen, F. S. (2000b). Effect of short-term hyperventilation on cerebral blood flow autoregulation in patients with acute bacterial meningitis. Stroke 2000. 31, 5, 1116-1122.
- Møller, K., Strauss, G. I., Qvist, J., Fonsmark, L., Knudsen, G. M., Larsen, F. S., Krabbe, K. S., Skinhøj, P., and Pedersen, B. K. (2002). Cerebral blood flow and oxidative metabolism during human endotoxemia. J Cereb.Blood Flow Metab 22, 10, 1262-1270.
- Møller, K., Tofteng, F., Qvist, T., Sahl, C., Sønderkaer, S., and Pedersen, B. K. (2005). Cerebral output of cytokines in patients with pneumococcal meningitis. Crit Care Med 33, 5, 979-983.
- Molyneux, E. M., Walsh, A. L., Forsyth, H., Tembo, M., Mwenechanya, J., Kayira, K., Bwanaisa, L., Njobvu, A., Rogerson, S., and Malenga, G. (2002). Dexamethasone treatment in childhood bacterial meningitis in Malawi: a randomised controlled trial. Lancet 360, 9328, 211-218.
- Mustafa, M. M., Lebel, M. H., Ramilo, O., Olsen, K. D., Reisch, J. S., Beutler, B., and McCracken, G. H., Jr. (1989). Correlation of interleukin-1 beta and cachectin concentrations in cerebrospinal fluid and outcome from bacterial meningitis. J.Pediatr. 115, 2, 208-213.
- Nau, R. (2000). Osmotherapy for elevated intracranial pressure A critical reappraisal. Clin.Pharmacokinet. 38, 1, 23-40.
- Nau, R. and Bruck, W. (2002). Neuronal injury in bacterial meningitis: mechanisms and implications for therapy. Trends Neurosci. 25, 1, 38-45.
- Nau, R., Sorgel, F., and Prange, H. W. (1994). Lipophilicity at pH 7.4 and molecular size govern the entry of the free serum fraction of drugs into the cerebrospinal fluid in humans with uninflamed meninges. J.Neurol Sci. 122, 1, 61-65.
- Nau, R., Soto, A., and Bruck, W. (1999a). Apoptosis of neurons in the dentate gyrus in humans suffering from bacterial meningitis. J Neuropathol Exp Neurol 58, 265-274.
- Nau, R., Wellmer, A., Soto, A., Koch, K., Schneider, O., Schmidt, H., Gerber, J., Michel, U., and Bruck, W. (1999b). Rifampin reduces early mortality in experimental Streptococcus pneumoniae meningitis. J Infect Dis 179, 1557-1560.
- Netter (1887). De la meningite due pneumocoque (avec ou sans pneumonie). Archives Générales de Médicine 19, 257-277.
- Nielsen, S. V. and Henrichsen, J. (1993). Capsular types and susceptibility to penicillin of pneumococci isolated from cerebrospinal fluid or blood in Denmark, 1983-1988. Scand.J.Infect.Dis. 25, 165-170.
- Denmark, 1983-1988. Scand.J.Infect.Dis. 25, 165-170.
 Nockher, W. A., Wick, M., and Pfister, H. W. (1999). Cerebrospinal fluid levels of soluble CD14 in inflammatory and non-inflammatory diseases of the CNS: upregulation during bacterial infections and viral meningitis. J.Neuroimmunol. 101, 2, 161-169.
- O'Reilly, T., Andes, D., Østergaard, C., and Frimodt-Møller, N. (2005). Evalution of antimicrobials in experimental animal infections. In Antibiotics in laboratory medicine, 5. edition.
- O'Reilly, T., Østergaard, C., Vaxelaire, J., and Zak, O. (2007). Systemic inflammation alters the inflammatory response in experimental LPS-induced meningitis. Clin Exp Immunol 147, 1, 112-119.
- Oates-Whitehead, R. M., Maconochie, I., Baumer, H., and Stewart, M. E. (2005). Fluid therapy for acute bacterial meningitis. Cochrane.Database.Syst.Rev.3, CD004786-
- Ohga, S., Aoki, T., Okada, K., Akeda, H., Fujioka, K., Ohshima, A., Mori, T., Minamishima, I., and Ueda, K. (1994). Cerebrospinal fluid concentrations of interleukin-1 beta, tumour necrosis factor-alpha, and interferon gamma in bacterial meningitis. Arch.Dis.Child 70, 123-125.
- Orihuela, C. J., Gao, G., Francis, K. P., Yu, J., and Tuomanen, E. I. (2004a). Tissue-specific contributions of pneumococcal virulence factors to pathogenesis. J Infect Dis 190, 9, 1661-1669.
- Orihuela, C. J., Gao, G., McGee, M., Yu, J., Francis, K. P., and Tuomanen, E. (2003). Organ-specific models of Streptococcus pneumoniae disease. Scand.J Infect.Dis. 35, 9, 647-652.
- Orihuela, C. J., Radin, J. N., Sublett, J. E., Gao, G., Kaushal, D., and Tuomanen, E. I. (2004b). Microarray Analysis of Pneumococcal Gene Expression during Invasive Disease. Infect Immun. 72, 10, 5582-5596.
- Ossege, L. M., Voss, B., Wiethege, T., Sindern, E., and Malin, J. P. (1994). Detection of transforming growth factor beta 1 mRNA in cerebrospinal fluid cells of patients with meningitis by non- radioactive in situ hybridization. J.Neurol. 242, 1, 14-19.
- Østergaard, C. (2000). The inflammatory respons in bacterial meningitis. An Experimental meningitis model. Ph.d.Dissertation
- Østergaard, C., Benfield, T., Frimodt-Møller, N., Espersen, F., Larsen, C. G., Mukaida, N., Matsushima, K., and Lundgren, J. D. (2000a). Treatment with a monoclonal antibody to interleukin-8 attenuates the pleocytosis in experimental pneumococcal meningitis in rabbits when given intravenously, but not intracisternally. Clin.Exp.Immunol. 122, 207-211.
- Østergaard, C., Benfield, T., Gesser, B., Kharazmi, A., Frimodt-Møller, N., Espersen, F., and Lundgren, J. D. (1999). Pretreatment with granulocyte colony-stimulating factor attenuates the inflammatory response but not

the bacterial load in cerebrospinal fluid during experimental pneumococcal meningitis in rabbits. Infection and Immunity 67, 7, 3430-3436.

- Østergaard, Č., Benfield, T., and Lundgren, J. D. (2002a). Macrophage Migration Inhibitory Factor is elevated in Cerebrospinal Fluid from Patients with Bacterial Meningitis. Abstract P570, 12th European Congress of Clinical Microbiology and Infectious Diseases, Milano
- Østergaard, C., Benfield, T., Lundgren, J. D., and Eugen-Olsen, J. (2004a). Soluble urokinase receptor is elevated in cerebrospinal fluid from patients with purulent meningitis and is associated with fatal outcome. Scand.J.Infect.Dis. 36, 1, 14-19.
- Østergaard, C., Benfield, T. L., Sellebjerg, F., Kronborg, G., Lohse, N., and Lundgren, J. D. (1996). Interleukin-8 in cerebrospinal fluid from patients with septic and aseptic meningitis. Eur.J.Clin.Microbiol.Infect.Dis. 15, 166-169.
- Østergaard, C., Brandt, C., Konradsen, H. B., and Samuelsson, S. (2004b). Differences in survival, brain damage and CSF cytokine kineticts due to meningitis caused by three different Streptococcus pneumoniae serotypes. Evaluation in humans and in two experimental meningitis models. J.Infec.Dis. 190, 7, 1212-1220.
- Østergaard, C., Høiby, N., Konradsen, H. B., and Samuelsson, S. (2006a). Prehospital diagnostic and therapeutic management of otogenic Streptococcus pneumoniae meningitis. Scand.J.Infect.Dis. 38, 172-180.
- Østergaard, C., Johansen, J. S., Benfield, T., Price, P. A., and Lundgren, J. D. (2002b). YKL-40 is elevated in cerebrospinal fluid from patients with purulent meningitis. Clin.Diagn.Lab Immunol. 9, 3, 598-604.
- Østergaard, C., Konradsen, H. B., and Samuelsson, S. (2005). Clinical presentation and prognostic factors of Streptococcus pneumoniae meningitis according the focus of infection. BMC.Infect Dis 5, 93, 1-11.
- Østergaard, C., O'Reilly, T., Brandt, C., Frimodt-Møller, N., and Lundgren, J. D. (2006b). The influence of the blood bacterial load on the meningeal inflammatory response in Streptococcus pneumoniae meningitis. BMC.Infect Dis 6, 78, 1-7.
- Østergaard, C., Sørensen, T. K., Knudsen, J. D., and Frimodt-Møller, N. (1998). Evaluation of moxifloxacin, a new 8-methoxyquinolone, for treatment of meningitis caused by a penicillin-resistant pneumococcus in rabbits. Antimicrobial Agents and Chemotherapy 42, 7, 1706-1712.
- Østergaard, C., Yieng-Kow, R. V., Benfield, T., Frimodt-Møller, N., Espersen, F., and Lundgren, J. D. (2000b). Inhibition of leukocyte entry into the brain by the selectin-blocker, fucoidin decreases interleukin (IL)-1 levels, but increases IL-8 levels in cerebrospinal fluid during experimental pneumococcal meningitis in rabbits. Infection and Immunity 68, 6, 3153-3157.
- Østergaard, C., Yieng-Kow, R. V., Knudsen, J. D., Frimodt-Møller, N., and Espersen, F. (2003). Evaluation of fusidic acid in therapy of experimental Staphylococcus aureus meningitis. J.Antimicrob.Chemother 51, 5, 1301-1305.
- Paris, M. M., Hickey, S. M., Trujillo, M., Ahmed, A., Olsen, K., and Mc-Cracken, G. H., Jr. (1997). The effect of interleukin-10 on meningeal inflammation in experimental bacterial meningitis. J Infect Dis 176, 5, 1239-1246.
- Paul, R., Angele, B., Sporer, B., Pfister, H. W., and Koedel, U. (2004). Inflammatory response during bacterial meningitis is unchanged in Fas- and Fas ligand-deficient mice. J Neuroimmunol. 152, 1-2, 78-82.
- Paul, R., Koedel, U., and Pfister, H. W. (2003a). Using knockout mice to study experimental meningitis. Arch.Immunol.Ther.Exp.(Warsz.) 51, 5, 315-326.
- Paul, R., Koedel, U., Winkler, F., Kieseier, B. C., Fontana, A., Kopf, M., Hartung, H. P., and Pfister, H. W. (2003b). Lack of IL-6 augments inflammatory response but decreases vascular permeability in bacterial meningitis. Brain 126, Pt 8, 1873-1882.
- Paul, R., Lorenzl, S., Koedel, U., Sporer, B., Vogel, U., Frosch, M., and Pfister, H. W. (1998). Matrix metalloproteinases contribute to the blood-brain barrier disruption during bacterial meningitis. Ann Neurol. 44, 4, 592-600.
- Paul, R., Winkler, F., Bayerlein, I., Popp, B., Pfister, H. W., and Koedel, U. (2005). Urokinase-type plasminogen activator receptor regulates leukocyte recruitment during experimental pneumococcal meningitis. J Infect Dis 191, 5, 776-782.
- Paulson, O. B., Brodersen, P., Hansen, E. L., and Kristensen, H. S. (1974). Regional cerebral blood flow, cerebral metabolic rate of oxygen, and cerebrospinal fluid acid-base variables in patients with acute meningitis and with acute encephalitis. Acta Med.Scand. 196, 3, 191-198.
- Pedersen, F. K. and Henrichsen, J. (1983). Pneumococcal meningitis and bacteraemia in Danish children 1969-1978. Serotypes, incidence and outcome. Acta Pathol.Microbiol.Immunol.Scand. [B] 91, 2, 129-134.
- Pedersen, M., Brandt, C. T., Knudsen, G. M., Østergaard, C., Skinhøj, P., Frimodt-Møller, N., and Møller, K. (2007). Cerbral blood flow autoregulation in early experimental Streptococcus pneumoniae meningitis. J.Appl.Physiol. 102, 1, 72-78.
- Pfister, H. W., Fontana, A., Tauber, M. G., Tomasz, A., and Scheld, W. M. (1994). Mechanisms of brain injury in bacterial meningitis: workshop summary. Clin.Infect.Dis. 19, 463-479.
- Pfister, H. W., Frei, K., Ottnad, B., Koedel, U., Tomasz, A., and Fontana, A.

(1992a). Transforming growth factor beta 2 inhibits cerebrovascular changes and brain edema formation in the tumor necrosis factor alphaindependent early phase of experimental pneumococcal meningitis. J Exp.Med. 176, 265-268.

- Pfister, H. W., Koedel, U., Dirnagl, U., Haberl, R. L., Feiden, W., and Einhaupl, K. M. (1990a). Superoxide dismutase inhibits brain oedema formation in experimental pneumococcal meningitis. Acta Neurochir. Suppl. Wien. 51, 378-380.
- Pfister, H. W., Koedel, U., Dirnagl, U., Haberl, R. L., Ruckdeschel, G., and Einhäupl, K. M. (1992b). Effect of catalase on regional cerebral blood flow and brain edema during the early phase of experimental pneumococcal meningitis. J.Infect.Dis. 166, 6, 1442-1445.
- Pfister, H. W., Koedel, U., Haberl, R. L., Dirnagl, U., Feiden, W., Ruckdeschel, G., and Einhaupl, K. M. (1990b). Microvascular changes during the early phase of experimental bacterial meningitis. J Cereb.Blood Flow Metab. 10, 914-922.
- Pfister, H. W., Koedel, U., Lorenzl, S., and Tomasz, A. (1992c). Antioxidants attenuate microvascular changes in the early phase of experimental pneumococcal meningitis in rats. Stroke 23, 1798-1804.
- Pfister, L. A., Tureen, J. H., Shaw, S., Christen, S., Ferriero, D. M., Tauber, M. G., and Leib, S. L. (2000). Endothelin inhibition improves cerebral blood flow and is neuroprotective in pneumococcal meningitis. Ann.Neurol. 47, 3, 329-335.
- Polissi, A., Pontiggia, A., Feger, G., Altieri, M., Mottl, H., Ferrari, L., and Simon, D. (1998). Large-scale identification of virulence genes from Streptococcus pneumoniae. Infection and Immunity 66, 12, 5620-5629.
- Poulsen, K., Reinholdt, J., Jespersgaard, C., Boye, K., Brown, T. A., Hauge, M., and Kilian, M. (1998). A comprehensive genetic study of streptococcal immunoglobulin A1 proteases: evidence for recombination within and between species. Infection and Immunity 66, 1, 181-190.
- Pracht, D., Elm, C., Gerber, J., Bergmann, S., Rohde, M., Seiler, M., Kim, K. S., Jenkinson, H. F., Nau, R., and Hammerschmidt, S. (2005). PavA of Streptococcus pneumoniae Modulates Adherence, Invasion, and Meningeal Inflammation. Infect Immun. 73, 5, 2680-2689.
- Prasad, K., Singhal, T., Jain, N., and Gupta, P. K. (2004). Third generation cephalosporins versus conventional antibiotics for treating acute bacterial meningitis. Cochrane.Database.Syst.Rev.2, CD001832-
- Prymula, R., Peeters, P., Chrobok, V., Kriz, P., Novakova, E., Kaliskova, E., Kohl, I., Lommel, P., Poolman, J., Prieels, J. P., and Schuerman, L. (2006). Pneumococcal capsular polysaccharides conjugated to protein D for prevention of acute otitis media caused by both Streptococcus pneumoniae and non-typable Haemophilus influenzae: a randomised double-blind efficacy study. Lancet 367, 9512, 740-748.
- Quade, F. and Kristensen, K. P. (1962). Purulent meningitis. A review of 658 cases. Acta Med.Scand. 171, 543-
- Quagliarello, V. J., Long, W. J., and Scheld, W. M. (1986). Morphologic alterations of the blood-brain barrier with experimental meningitis in the rat. Temporal sequence and role of encapsulation. J.Clin.Invest. 77, 4, 1084-1095.
- Quagliarello, V. J., Wispelwey, B., Long, W. J., Jr., and Scheld, W. M. (1991). Recombinant human interleukin-1 induces meningitis and blood- brain barrier injury in the rat. Characterization and comparison with tumor necrosis factor. J.Clin.Invest. 87, 4, 1360-1366.
- Ramilo, O., Mustafa, M. M., Porter, J., Saez Llorens, X., Mertsola, J., Olsen, K. D., Luby, J. P., Beutler, B., and McCracken, G. H., Jr. (1990a). Detection of interleukin 1 beta but not tumor necrosis factor- alpha in cerebrospinal fluid of children with aseptic meningitis. Am.J.Dis.Child. 144, 3, 349-352.
- Ramilo, O., Saez Llorens, X., Mertsola, J., Jafari, H., Olsen, K. D., Hansen, E. J., Yoshinaga, M., Ohkawara, S., Nariuchi, H., and McCracken, G. H., Jr. (1990b). Tumor necrosis factor alpha/cachectin and interleukin 1 beta initiate meningeal inflammation. J.Exp.Med. 172, 2, 497-507.
- Ratilal, B., Costa, J., and Sampaio, C. (2006). Antibiotic prophylaxis for preventing meningitis in patients with basilar skull fractures. Cochrane.Database.Syst.Rev.1, CD004884-
- Biernath, K. R., Broder, K., Manning, S., Avashia, S., Victor, M., Costa, P., Devine, O., Graham, A., and Boyle, C. (2003). Risk of bacterial meningitis in children with cochlear implants. N Engl J Med 349, 5, 435-445.
- Rendi-Wagner, P., Georgopoulos, A., Kundi, M., Mutz, I., Mattauch, M., Nowak, J., Mikolasek, A., Vecsei, A., and Kollaritsch, H. (2004). Prospective surveillance of incidence, serotypes and antimicrobial susceptibility of invasive Streptococcus pneumoniae among hospitalized children in Austria. J Antimicrob.Chemother. 53, 5, 826-831.
- Rieckmann, P., Albrecht, M., Ehrenreich, H., Weber, T., and Michel, U. (1995). Semi-quantitative analysis of cytokine gene expression in blood and cerebrospinal fluid cells by reverse transcriptase polymerase chain reaction. Research in Experimental Medicine 195, 17-29.
- Ring, A., Weiser, J. N., and Tuomanen, E. I. (1998). Pneumococcal trafficking across the blood-brain barrier – Molecular analysis of a novel bidirectional pathway. J Clin.Invest. 102, 2, 347-360.
- Roine, I., Foncea, L. M., Cofre, J., Ledermann, W., and Peltola, H. (1992). Serum C-reactive protein vs. tumor necrosis factor alpha and interleukin

1 beta of the cerebrospinal fluid in diagnosis of bacterial meningitis with low cerebrospinal fluid cell count. Pediatr.Infect.Dis.J. 11, 1057-1058.

- Rozdzinski, E., Jones, T., Burnette, W. N., Burroughs, M., and Tuomanen, E. (1993). Antiinflammatory effects in experimental meningitis of prokaryotic peptides that mimic selectins. J.Infect.Dis. 168, 6, 1422-1428.
- Rozdziński, E., Sandros, J., van-der-Flier, M., Young, A., Spellerberg, B., Bhattacharyya, C., Straub, J., Musso, G., Putney, S., Starzyk, R., and et-al (1995a). Inhibition of leukocyte-endothelial cell interactions and inflammation by peptides from a bacterial adhesin which mimic coagulation factor X. J.Clin.Invest. 95, 1078-1085.
- Rozdzinski, E., Spellerberg, B., van der Flier, M., Bhattacharyya, C., Hoepelman, A. I., Moran, M. A., Jarpe, A., Putney, S. D., Starzyk, R. M., and Tuomanen, E. (1995b). Peptide from a prokaryotic adhesin blocks leukocyte migration in vitro and in vivo. J.Infect.Dis. 172, 3, 785-793.
- Saez-Llorens, X., McCoig, C., Feris, J. M., Vargas, S. L., Klugman, K. P., Hussey, G. D., Frenck, R. W., Falleiros-Carvalho, L. H., Arguedas, A. G., Bradley, J., Arrieta, A. C., Wald, E. R., Pancorbo, S., McCracken, G. H., Jr., and Marques, S. R. (2002). Quinolone treatment for pediatric bacterial meningitis: a comparative study of trovafloxacin and ceftriaxone with or without vancomycin. Pediatr.Infect.Dis.J 21, 1, 14-22.
- Sandros, J., Rozdzinski, E., and Tuomanen, E. (1994). Peptides from pertussis toxin interfere with neutrophil adherence in vitro and counteract inflammation in vivo. Microb.Pathog. 16, 3, 213-220.
 Saukkonen, K., Sande, S., Cioffe, C., Wolpe, S., Sherry, B., Cerami, A., and
- Saukkonen, K., Sande, S., Cioffe, C., Wolpe, S., Sherry, B., Cerami, A., and Tuomanen, E. (1990). The role of cytokines in the generation of inflammation and tissue damage in experimental gram-positive meningitis. J.Exp.Med. 171, 439-448.
- Schaad, U. B., Suter, S., Gianella Borradori, A., Pfenninger, J., Auckenthaler, R., Bernath, O., Cheseaux, J. J., and Wedgwood, J. (1990). A comparison of ceftriaxone and cefuroxime for the treatment of bacterial meningitis in children. N.Engl.J.Med. 322, 3, 141-147.
- Schaper, M., Leib, S. L., Meli, D. N., Brandes, R. P., Tauber, M. G., and Christen, S. (2003). Differential effect of p47phox and gp91phox deficiency on the course of Pneumococcal Meningitis. Infection and Immunity 71, 7, 4087-4092.
- Scheld, W. M., Dacey, R. G., Winn, H. R., Welsh, J. E., Jane, J. A., and Sande, M. A. (1980). Cerebrospinal fluid outflow resistance in rabbits with experimental meningitis. Alterations with penicillin and methylprednisolone. J Clin.Invest. 66, 2, 243-253.
- Schneider, O., Michel, U., Zysk, G., Dubuis, O., and Nau, R. (1999). Clinical outcome in pneumococcal meningitis correlates with CSF lipoteichoic acid concentrations. Neurology 53, 7, 1584-1587.
- Schnell, L., Fearn, S., Schwab, M. E., Perry, V. H., and Anthony, D. C. (1999). Cytokine-induced acute inflammation in the brain and spinal cord. J Neuropathol Exp Neurol 58, 245-254.
- Schwarz, S., Bertram, M., Schwab, S., Andrassy, K., and Hacke, W. (2000). Serum procalcitonin levels in bacterial and abacterial meningitis. Crit.Care Med. 28, 6, 1828-1832.
- Simberkoff, M. S., Moldover, N. H., and Rahal, J., Jr. (1980). Absence of detectable bactericidal and opsonic activities in normal and infected human cerebrospinal fluids. A regional host defense deficiency. J.Lab.Clin.Med. 95, 3, 362-372.
- Smith, W. B., Gamble, J. R., Clark Lewis, I., and Vadas, M. A. (1991). Interleukin-8 induces neutrophil transendothelial migration. Immunology 72, 1, 65-72.
- Sørensen, U. B. (1995). Pneumococcal polysaccharide antigens: capsules and C-polysaccharide. An immunochemical study. Dan.Med.Bull. 42, 1, 47-53.
- Southard, E. E. and Keene, C. W. (1906). A study of brain infections with the pneumococcus. JAMA 46, 13-21.
- Spranger, M., Schwab, S., Krempien, S., Winterholler, M., Steiner, T., and Hacke, W. (1996). Excess glutamate levels in the cerebrospinal fluid predict clinical outcome of bacterial meningitis. Arch.Neurol. 53, 10, 992-996.
- Spreer, A., Kerstan, H., Bottcher, T., Gerber, J., Siemer, A., Zysk, G., Mitchell, T. J., Eiffert, H., and Nau, R. (2003). Reduced Release of Pneumolysin by Streptococcus pneumoniae In Vitro and In Vivo after Treatment with Nonbacteriolytic Antibiotics in Comparison to Ceftriaxone. Antimicrobial Agents and Chemotherapy 47, 8, 2649-2654.
- Sprenger, H., Rosler, A., Tonn, P., Braune, H. J., Huffmann, G., and Gemsa, D. (1996). Chemokines in the cerebrospinal fluid of patients with meningitis. Clin Immunol Immunopathol 80, 2, 155-161.
- Stearman, M. and Southgate, H. J. (1994). The use of cytokine and C-reactive protein measurements in cerebrospinal fluid during acute infective meningitis. Ann.Clin.Biochem. 31, 255-261.
- Stephens, R. W., Nielsen, H. J., Christensen, I. J., Thorlacius-Ussing, O., Sorensen, S., Dano, K., and Brunner, N. (1999). Plasma urokinase receptor levels in patients with colorectal cancer: relationship to prognosis. J Natl.Cancer Inst. 91, 10, 869-874.
- Stewart, F. W. (1927). Local specific therapy of experimental pneumococcal meningitis. I. Experimental pneumococcal meningitis in rabbits. J.Exp.Med. 46, 391-407.

- Stuertz, K., Schmidt, H., Trostdorf, F., Eiffert, H., Mader, M., and Nau, R. (1999). Lower lipoteichoic and teichoic acid CSF concentrations during treatment of pneumococcal meningitis with non- bacteriolytic antibiotics than with ceftriaxone. Scand.J.Infec.Dis. 31, 4, 367-370.
- Swartz, M. N. (2004). Bacterial meningitis--a view of the past 90 years. N.Engl.J Med. 351, 18, 1826-1828.
- Takala, A. K., Jero, J., Kela, E., Ronnberg, P. R., Koskenniemi, E., and Eskola, J. (1995). Risk factors for primary invasive pneumococcal disease among children in Finland. JAMA 273, 11, 859-864.
 Tang, T., Frenette, P. S., Hynes, R. O., Wagner, D. D., and Mayadas, T. N.
- Tang, T., Frenette, P. S., Hynes, R. O., Wagner, D. D., and Mayadas, T. N. (1996). Cytokine-induced Meningitis Is Dramatically Attenuated in Mice Deficient in Endothelial Selectins. J.Clin.Invest. 97, 2485-2490.
- Tauber, M. G., Borschberg, U., and Sande, M. A. (1988). Influence of granulocytes on brain edema, intracranial pressure, and cerebrospinal fluid concentrations of lactate and protein in experimental meningitis. J.Infect.Dis. 157, 456-464.
- Tauber, M. G., Burroughs, M., Niemoller, U. M., Kuster, H., Borschberg, U., and Tuomanen, E. (1991). Differences of pathophysiology in experimental meningitis caused by three strains of Streptococcus pneumoniae. J.Infect.Dis. 163, 806-811.
- Tauber, M. G., Doroshow, C. A., Hackbarth, C. J., Rusnak, M. G., Drake, T. A., and Sande, M. A. (1984). Antibacterial activity of beta-lactam antibiotics in experimental meningitis due to Streptococcus pneumoniae. J.Infect.Dis. 149, 568-574.
- Tauber, M. G., Kennedy, S. L., Tureen, J. H., and Lowenstein, D. H. (1992). Experimental pneumococcal meningitis causes central nervous system pathology without inducing the 72-kd heat shock protein. Am.J.Pathol. 141, 53-60.
- Tauber, M. G., Khayam-Bashi, H., and Sande, M. A. (1985). Effects of ampicillin and corticosteroids on brain water content, cerebrospinal fluid pressure, and cerebrospinal fluid lactate levels in experimental pneumococcal meningitis. J.Infect.Dis. 151, 528-534.
- Tauber, M. G. and Moser, B. (1999). Cytokines and chemokines in meningeal inflammation: Biology and clinical implications. Clin.Infect.Dis. 28, 1-12.
- Temesvári, P., Abrahám, C. S., Gellén, J., Jr., Speer, C. P., Kovács, J., and Megyeri, P. (1995). Elastase given intracisternally opens blood-brain barrier in newborn piglets. Biol.Neonate. 67, 1, 59-63.
- The Research Committee of the BSSI (1995). Bacterial Meningitis: Causes For Concern. Jornal of Infection 30, 89-94.
- Tong, H. H., Blue, L. E., James, M. A., and DeMaria, T. F. (2000). Evaluation of the virulence of a Streptococcus pneumoniae neuraminidase-deficient mutant in nasopharyngeal colonization and development of otitis media in the chinchilla model. Infection and Immunity 68, 2, 921-924.
- Trostdorf, F., Bruck, W., SchmitzSalue, M., Stuertz, K., Hopkins, S. J., van Rooijen, N., Huitinga, I., and Nau, R. (1999). Reduction of meningeal macrophages does not decrease migration of granulocytes into the CSF and brain parenchyma in experimental pneumococcal meningitis. J.Neuroimmunol. 99, 2, 205-210.
- Tumani, H., Smirnov, A., Barchfeld, S., Olgemoller, U., Maier, K., Lange, P., Bruck, W., and Nau, R. (2000). Inhibition of glutamine synthetase in rabbit pneumococcal meningitis is associated with neuronal apoptosis in the dentate gyrus. Glia 30, 1, 11-18.
- Tunkel, A. R., Hartman, B. J., Kaplan, S. L., Kaufman, B. A., Roos, K. L., Scheld, W. M., and Whitley, R. J. (2004). Practice guidelines for the management of bacterial meningitis. Clin.Infect Dis 39, 9, 1267-1284.
- Tuomanen, E., Hengstler, B., Řich, R., Bray, M. A., Zak, O., and Tomasz, A. (1987). Nonsteroidal anti-inflammatory agents in the therapy for experimental pneumococcal meningitis. J.Infect.Dis. 155, 985-990.
- Tuomanen, E., Liu, H., Hengstler, B., Zak, O., and Tomasz, A. (1985a). The induction of meningeal inflammation by components of the pneumococcal cell wall. J.Infect.Dis. 151, 5, 859-868.
- Tuomanen, E., Pollack, H., Parkinson, A., Davidson, M., Facklam, R., Rich, R., and Zak, O. (1988). Microbiological and clinical significance of a new property of defective lysis in clinical strains of pneumococci. J.Infect.Dis. 158, 36-43.
- Tuomanen, E., Tomasz, A., Hengstler, B., and Zak, O. (1985b). The relative role of bacterial cell wall and capsule in the induction of inflammation in pneumococcal meningitis. J.Infect.Dis. 151, 535-540.
- Tuomanen, E. I., Saukkonen, K., Sande, S., Cioffe, C., and Wright, S. D. (1989). Reduction of inflammation, tissue damage, and mortality in bacterial meningitis in rabbits treated with monoclonal antibodies against adhesion-promoting receptors of leukocytes. J.Exp.Med. 170, 3, 959-969.
- Tureen, J. H., Dworkin, R. J., Kennedy, S. L., Sachdeva, M., and Sande, M. A. (1990). Loss of cerebrovascular autoregulation in experimental meningitis in rabbits. J.Clin.Invest. 85, 2, 577-581.
- Tureen, J. H., Tauber, M. G., and Sande, M. A. (1991). Effect of indomethacin on the pathophysiology of experimental meningitis in rabbits. J.Infect.Dis. 163, 647-649.
- Tureen, J. H., Tauber, M. G., and Sande, M. A. (1992). Effect of hydration status on cerebral blood flow and cerebrospinal fluid lactic acidosis in rabbits with experimental meningitis. J.Clin.Invest. 89, 3, 947-953.
- van de Beek, D. and de Gans, J. (2004). Dexamethasone and pneumococcal meningitis. Ann.Intern.Med. 141, 4, 327.

- van de Beek, D., de Gans, J., McIntyre, P., and Prasad, K. (2003). Corticosteroids in acute bacterial meningitis. Cochrane.Database.Syst.Rev.3, CD004405.
- van de Beek, D., de Gans, J., Spanjaard, L., Weisfelt, M., Reitsma, J. B., and Vermeulen, M. (2004). Clinical features and prognostic factors in adults with bacterial meningitis. N.Engl.J Med. 351, 18, 1849-1859.
- van de, Beek D., de Gans, J., Tunkel, A. R., and Wijdicks, E. F. (2006). Community-acquired bacterial meningitis in adults. N.Engl.J.Med. 354, 1, 44-53.
- Viladrich, P. F., Gudiol, F., Liñares, J., Pallarés, R., Sabaté, I., Rufi, G., and Ariza, J. (1991). Evaluation of vancomycin for therapy of adult pneumococcal meningitis. Antimicrob.Agents Chemother. 35, 12, 2467-2472.
- Waage, A., Halstensen, A., Shalby, R., Brandtzaeg, P., Kierulf, P., and Espevik, T. (1989). Local production of tumor necrosis factor-alpha, interleukin 1, and interleukin 6 in meningococcal meningitis. J.Exp.Med. 170, 1859-1867.
- Weber, J. R., Angstwurm, K., Burger, W., Einhaupl, K. M., and Dirnagl, U. (1995). Anti ICAM-1 (CD 54) monoclonal antibody reduces inflammatory changes in experimental bacterial meningitis. J.Neuroimmunol. 63, 63-68.
- Weber, J. R., Freyer, D., Alexander, C., Schroder, N. W., Reiss, A., Kuster, C., Pfeil, D., Tuomanen, E. I., and Schumann, R. R. (2003). Recognition of pneumococcal peptidoglycan: an expanded, pivotal role for LPS binding protein. Immunity. 19, 2, 269-279.
- Weisfelt, M., van de, Beek D., Spanjaard, L., Reitsma, J. B., and de Gans, J. (2006). Clinical features, complications, and outcome in adults with pneumococcal meningitis: a prospective case series. Lancet Neurol. 5, 2, 123-129.
- Weiss, W., Figueroa, W., Shapino, W. H., and Flippin, H. F. (1967). Prognostic factors in pneumococcal meningitis. Arch.Intern.Med. 120, 517-524.
- Wellmer, A., Gerber, J., Ragheb, J., Zysk, G., Kunst, T., Smirnov, A., Bruck, W., and Nau, R. (2001). Effect of deficiency of tumor necrosis factor alpha or both of its receptors on Streptococcus pneumoniae central nervous system infection and peritonitis. Infection and Immunity 69, 11, 6881-6886.
- Wellmer, A., von Mering, M., Spreer, A., Diem, R., Eiffert, H., Noeske, C., Bunkowski, S., Gold, R., and Nau, R. (2004). Experimental pneumococcal meningitis: Impaired clearance of bacteria from the blood due to increased apoptosis in the spleen in Bcl-2-deficient mice. Infection and Immunity 72, 6, 3113-3119.
- Wellmer, A., Zysk, G., Gerber, J., Kunst, T., von Mering, M., Bunkowski, S., Eiffert, H., and Nau, R. (2002). Decreased Virulence of a Pneumolysin-Deficient Strain of Streptococcus pneumoniae in Murine Meningitis. Infection and Immunity 70, 11, 6504-6508.
- Whitney, C. G., Farley, M. M., Hadler, J., Harrison, L. H., Lexau, C., Reingold, A., Lefkowitz, L., Cieslak, P. R., Cetron, M., Zell, E. R., Jorgensen, J. H., and Schuchat, A. (2000). Increasing prevalence of multidrug-resistant Streptococcus pneumoniae in the United States. N.Engl.J.Med. 343, 26, 1917-1924.
- Whittle, H. C. and Greenwood, B. M. (1977). Cerebrospinal fluid immunoglobulins and complement in meningococcal meningitis. J Clin.Pathol. 30, 8, 720-722.
- Winkler, F., Koedel, U., Kastenbauer, S., and Pfister, H. W. (15-6-2001). Differential expression of nitric oxide synthases in bacterial meningitis: role of the inducible isoform for blood-brain barrier breakdown. J.Infect.Dis. 183, 12, 1749-1759.
- Wittenhagen, P., Kronborg, G., Weis, N., Nielsen, H., Obel, N., Pedersen, S. S., and Eugen-Olsen, J. (2004). The Plasma Level of Soluble Urokinase Plasminogen Activator Receptor is Elevated in Patients with Streptococcus pneumoniae Bacteremia and Carries Strong Prognostic Value. Clin. Microbiol.Infect. 10, 5, 409-415.
- Zhang, J. R., Mostov, K. E., Lamm, M. E., Nanno, M., Shimida, S., Ohwaki, M., and Tuomanen, E. (2000). The polymeric immunoglobulin receptor translocates pneumococci across human nasopharyngeal epithelial cells. Cell 102, 827-837.
- Zwahlen, A., Nydegger, U. E., Vaudaux, P., Lambert, P. H., and Waldvogel, F. A. (1982). Complement-mediated opsonic activity in normal and infected human cerebrospinal fluid: early response during bacterial meningitis. J.Infect.Dis. 145, 5, 635-646.
- Zwijnenburg, P. J., Polfliet, M. M., Florquin, S., van den Berg, T. K., Dijkstra, C. D., van Deventer, S. J., Roord, J. J., van der, Poll T., and van Furth, A. M. (2003a). CXC-chemokines KC and macrophage inflammatory protein-2 (MIP-2) synergistically induce leukocyte recruitment to the central nervous system in rats. Immunol.Lett. 85, 1, 1-4.
- Zwijnenburg, P. J., van der, Poll T., Florquin, S., Akira, S., Takeda, K., Roord, J. J., and van Furth, A. M. (2003b). Interleukin-18 gene-deficient mice show enhanced defense and reduced inflammation during pneumococcal meningitis. J Neuroimmunol. 138, 1-2, 31-37.
- Zwijnenburg, P. J., van der, Poll T., Florquin, S., Roord, J. J., and van Furth, A. M. (2003c). IL-1 receptor type 1 gene-deficient mice demonstrate an impaired host defense against pneumococcal meningitis. J Immunol. 170, 9, 4724-4730.

Zwijnenburg, P. J., van der, Poll T., Florquin, S., Roord, J. J., and van Furth,

A. M. (2003d). Interleukin-10 negatively regulates local cytokine and chemokine production but does not influence antibacterial host defense during murine pneumococcal meningitis. Infection and Immunity 71, 4, 2276-2279.

- Zwijnenburg, P. J., van der, Poll T., Florquin, S., van Deventer, S. J., Roord, J. J., and van Furth, A. M. (2001). Experimental pneumococcal meningitis in mice: a model of intranasal infection. J.Infect.Dis. 183, 7, 1143-1146.
- Zwijnenburg, P. J., van der, Poll T., Roord, J. J., and van Furth, A. M. (2006). Chemotactic factors in cerebrospinal fluid during bacterial meningitis. Infection and Immunity 74, 3, 1445-1451.
- Zysk, G., Brück, W., Fischer, F. R., Mäder, M., Rieckmann, P., and Nau, R. (1997a). Limited efficacy of pentoxifylline as anti-inflammatory agent in experimental pneumococcal meningitis. Clin.Exp Immunol 107, 3, 458-461.
- Zysk, G., Brück, W., Gerber, J., Brück, Y., Prange, H. W., and Nau, R. (1996). Anti-inflammatory treatment influences neuronal apoptotic cell death in the dentate gyrus in experimental pneumococcal meningitis. J Neuropathol Exp Neurol 55, 6, 722-728.
- Zysk, G., Brück, W., Huitinga, I., Fischer, F. R., Flachsbarth, F., van Rooijen, N., and Nau, R. (1997b). Elimination of blood-derived macrophages inhibits the release of interleukin-1 and the entry of leukocytes into the cerebrospinal fluid in experimental pneumococcal meningitis. J Neuroimmunol. 73, 1-2, 77-80.